

A DETAILED OVERVIEW OF EXTRA PULMONARY TUBERCULOSIS PATHOPHYSIOLOGY AND ITS DIAGNOSTIC METHODS

Talluri Rameshwari KR

Department of Microbiology, JSS Academy of Higher Education & Research, Sri
Shivarathreeshwara Nagar, Mysuru, India. _

Nazia Khan

Assistant Professor , Clinical Microbiology, Basic Medical Science, College Of Medicine,
Majmaah University, Al-Majmaah, Riyadh, Saudi Arabia. _

Shubhkarmanjit Singh Bawa

Senior lecturer , Department of Periodontology , Himachal Dental College, Sundernagar ,
Himachal Pradesh , _

Parul Sharma

Senior lecturer

Department of Periodontology , Himachal Dental College, Sundernagar. Himachal Pradesh

Vijayanandhan Venkatesan

Faculty of Pharmacognosy, JKKN College of Pharmacy, JKKN Educational Institutions,
Kumarapalayam, Namakkal, Tamil Nadu. , Orcid Id - 0000-0003-0584-6243

Ritik Kashwani

BDS, Private Practitioner, Ex-Junior Resident, School of Dental Sciences, Sharda
University, Greater Noida. Orcid Id - 0009-0008-8922-7522.

Sumana Kumar

Associate Professor, Department of Microbiology, JSS Academy of Higher Education &
Research, Sri Shivarathreeshwara Nagar, Mysuru, India, Orcid Id : 0000-0003-2482-439

Abstract

A hematogenous spread of *Mycobacterium tuberculosis*(MTB) in human body leading to Extra pulmonary tuberculosis (EPTB). About 25% of Tuberculosis (TB) cases involved EPTB by the spread through blood and lymphatic routes further outspread to other organs. EPTB most usually affects the lymphatic system. In the event of necrotic lymph nodes and other organ-specific imaging abnormalities, the likelihood of detecting EPTB increases. In cases of necrotic lymph nodes and other organ-specific imaging abnormalities, the likelihood of detecting EPTB increases. Immunosuppressed individuals exhibit the most pronounced central nervous system infection spread. The precise diagnosis might be challenging to make since clinical symptoms and imaging test results may be ambiguous. It is often

required to involve invasive diagnostic testing for the collected biological samples. The cultures are still gold standard methods as a stable detection of mycobacterial DNA despite the use of advance molecular methods as an early diagnosis. The EPTB treatment has 6 months antibiotic regimens similar to pulmonary tuberculosis. Further drug extension is advice solely in MTB affecting central nervous systems. The amount of microbial level and its position affect the susceptibility and accuracy of traditional examination techniques. It is usually necessary to gather source using invasive methods for histopathological and microbiology investigation.

Keywords: Extra pulmonary Tuberculosis, Pathophysiology, Lymphadenitis, Meningitis, Pleura, Diagnostics etc.,

1. Introduction

Tuberculosis caused by the bacteria *Mycobacterium tuberculosis* that results in a concerted interaction between biological and pathology procedures. The MTB has grown to flourish by invading the human being and staying in it for a considerable amount of duration manipulating the natural defences of its recipient. Each 18 to 24 hours, MTB, which is a mycolic acid-coated, not mobile internal infectious bacteria, multiplies its tissue [1, 2].

Over 1.7 to 2 billion individuals worldwide are affected with this disease, which results in over 4,000 deaths every day and 1.2 to 1.5 million deaths annually [1, 5]. Over 13 million Americans may have latent tuberculosis infections, according to estimates (LTBI). Active tuberculosis will develop in 5–10% of these LTBI patients [6]. Approximately fifteen million occurrences of persistent TB are identified each year, including India, South Africa, the Philippines, Indonesia, Bangladesh, Nigeria, Pakistan and China suffering the largest share of the incidence [4, 3, 7].

Patients with lower immunity are more likely to develop active TB. Human immunodeficiency virus (HIV) sufferers, recipients of transplants, those with insulin resistance, and people with silicosis are a few examples of cases. [7, 8].

Since individuals have been the bacteria's host for a very long time, the bacteria has developed unique antibacterial defences which enable it to remain within the host [1]. This article's goal is to compare all seven stages of persistent MTB disease, TB pathology, and disease transmission from a pharmacological perspective.

Aerosolization, macrophage phagocytosis, phagolysosome blockage and replication, T helper type 1 (TH1) response, granuloma formation, clinical signs, and transmission are the processes that are involved. Lesser intermediary processes [9]. In light of the molecular process, cellular motions and morphologies, and clinical symptoms, current research will examine these steps. The lungs, the most frequently damaged anatomical site, will be the subject of study. [7]. But in addition to the apparatus urogenitalis, and stomach structures, the illness has the ability to travel to the epidermis, nerves, his eyes. lymphocytes, connections, and skeletons.

The seven steps in MTB infection, tuberculosis pathogenesis, and disease transmission will be discussed first. These include transmission, granuloma formation, TH1 response, macrophage phagocytosis, phagolysosome blockage, clinical signs, and aerosolization [9,10]. The second goal is to outline the

pharmaceutical industry's perspective on tuberculosis and the possibilities for new treatments.

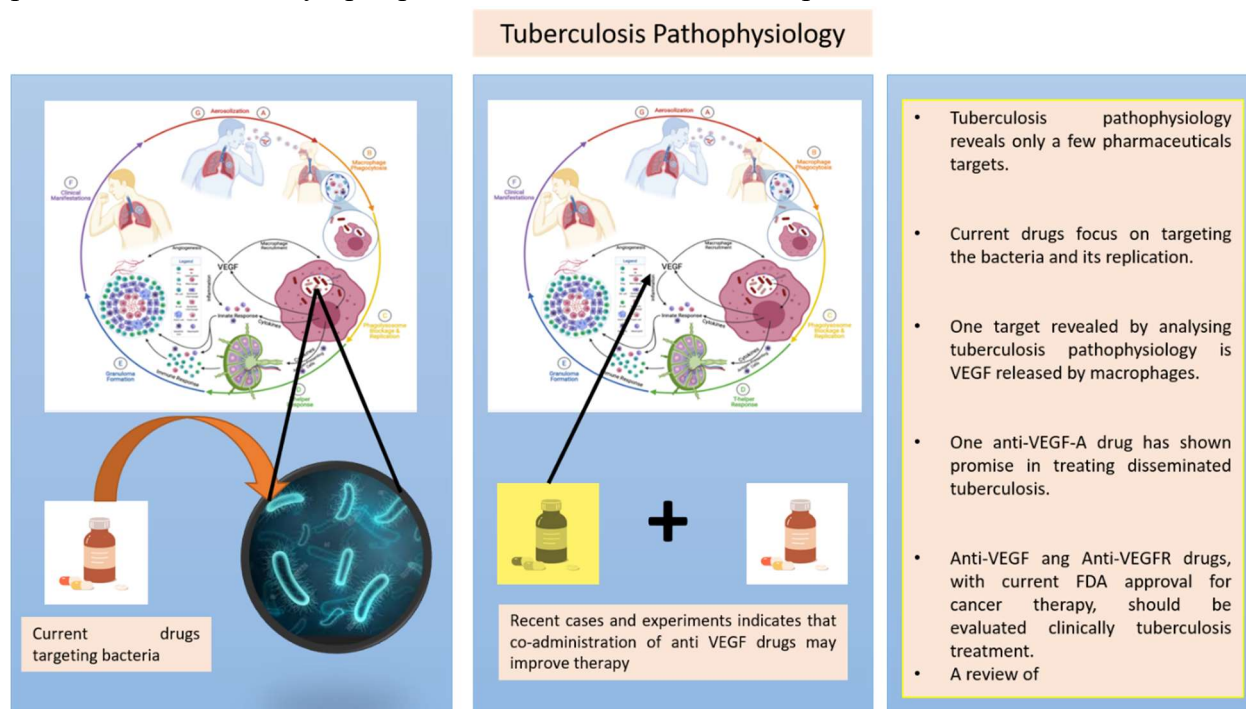


Figure 1: *Mycobacterium tuberculosis* pathophysiology

1. Tuberculous Lymphadenitis: Extrapulmonary tuberculosis (EPTB) is most usually seen as lymphadenitis. The most prevalent type of adenopathy is cervical adenopathy, but inguinal, axillary, mesenteric, mediastinal, and intermammary involvement have all been reported. [13, 17] While lymphadenitis was once thought to be a childhood disease, it now affects people in their 20s and 40s, and It most frequently affects women and immigrants in the United States[141]. Patients who do not have HIV usually have chronic, nontender lymphadenopathy. [15,17] Weight loss, nocturnal sweating, and fever are typical signs of HIV infection. [5,18] A dense mass of matted nodes eventually forms from the separate, rigid, and nontender nodes. If left untreated, *Mycobacterium tuberculosis* present on the fine-needle aspiration specimen increases test sensitivity. During anti-tuberculous therapy, affected nodes may expand or new nodes may appear, indicating an immune reaction to dead mycobacteria. HIV patients who begin antiretroviral medication concurrently have a similar outcome due to immune reconstitution. The removal of lymph nodes is typically not advised. When lymph nodes are swollen and prepared to empty, aspiration or incision and drainage seem to be successful treatments.

Pathophysiology: Lymphatic spread from the mediastinal lymph nodes results in cervical adenitis following the reactivation of a primary lung infection [6]. On the other hand, tuberculous lymphadenitis may be the initial symptom of a non-tuberculous mycobacteria infection. Young children are immediately exposed to it by bacilli in the oropharyngeal mucosa [5]. The risk of contracting tuberculosis outside of the lungs increases in the presence of necrotic lymph nodes. The germs can enter lymphatic vessels and spread by haemorrhaging into the sinus cavities, where latent cell lines can kill them. Inhaled microbes multiply in macrophages till the microglia' mediators and enzymes that break

down proteolytic material induce dying of cells, at which time the bacilli are ejected. A mass of cells is created by the activated macrophages and monocytes in order to stop getting infected and restrict the bacteria's ability to reproduce. During this stage, the granuloma eventually develops into an abscess with a decaying centre and surround granulated matter(**Fig. 1**) [6].

Imaging: There are no distinctive visualization characteristics of tuberculous adenitis. On the opposite hand, Reede et al. [7] have found four types of illness that are commonly observed on CT scans including (a) multidimensional enlargements of lymph nodes with heavy external development, (b) massive negligible-attenuation convergent a number in secondary growth and fatty plane damage, (c) uniform broad lymphatic system in addition to improvement, and (d) widened lymphatic fluid nodes along with slim-wall outskirts expansion and fatty tissue plane preserving it. The initially identified pair of patterns exhibit an ongoing transition from several decaying diseased lymph vessels to a solitary necrosis mass, enhancing the susceptibility of the TB testing procedure

2.Pleural Tuberculosis: A little more than five per cent. of instances of TB in the US are pulmonary. [19] Initial post-primary TB, persistent lung disease, and miliary tb are among situations that may end up in tuberculous effusions. Pleural TB often presents as a high temperature, breathing difficulties pleuritic pain in the chest, and a dry cough. Approximately twenty percent of the individuals have a small to big solitary pleural drainage visible on lung imaging, and 20% of participants have arterial abnormalities. 20 respiratory infections, lymphadenopathy, and depression that is not apparent on a chest x-ray On a chest CT scan, an MRI can be seen. Most of the time, there is a pulmonary enlargement that is wider than one centimetre.

The fluid inside the pleura is exudative and has a lymphoma majority (i.e., over fifty per cent of leukocyte in over ninety % of outflowing) [16,20,23]; among individuals with a shorter duration of indications, the pleural fluid is a first a stronghold of neutrophils. These could be observed.

The pleural fluid's pH and glucose levels may be low or normal. AFB smears of pleural fluid are infrequently 5% Positive unless the patient has tuberculous empyema. Pleural fluid cultures for *M. tuberculosis* are positive in fewer than 40% of patients. 20 More than 90% of the pleural biopsy specimen analysis's results are sensitive overall (caseating granulomas observation, AFB smear, and culture). 20 A tuberculin skin test is positive in two-thirds of cases. Biochemical indicators found in the pleural fluid, such as adenosine deaminase, interferon gamma, and lysozyme, can be useful.

Pathophysiology: Similarly to meningitis caused by tuberculosis, a large granulate discharge covers the cavity under the brain and irritates the cervical vertebrae and nerves

Exudate causes adhesions and fibrosis in chronic situations. Vasculitis, Lumbar TB, which also affects brain function, may end up in localised regional infarctions and swelling of your vertebral column. Infectious thrombosis of the spinal arteries can potentially cause ischemia [11,18].

3.Skeletal Tuberculosis: Up to 35% of instances of extrapulmonary tuberculosis are suspected to be caused by TB joints and bones. Skeletal TB most frequently affects the spine[25, 27].Following that are extraspinal tuberculous bone disease and weight-bearing joint tuberculous arthritic.

The most common location for TB (Pott's disease) is the thoracic spine. As the infection spreads to the

anteroinferior portion of the vertebral body, the intervertebral disc and surrounding vertebrae are destroyed. The characteristic radiographic appearance and palpable spinal protrusion (gibbus) are the result of anterior wedging and angulation of adjacent vertebral bodies with disc space obliteration. The paraspinal and psoas muscles can develop abscesses that spread to the surface or adjacent tissues. Patients with cord compression present with paraplegia, generalised discomfort, or both. A monoarthritis of the hip or knee, articular tuberculosis advances gradually. The indolent presentation is marked by pain, joint edoema, and limited range of motion. Abscesses and leaking sinuses are frequent in chronic conditions. Body-wide symptoms are usually absent. Nonspecific radiographic changes include soft tissue edoema, juxta-articular osteopenia, joint space narrowing, and subchondral erosions [26]. A particular type of extraspinal tuberculous osteomyelitis, which may impact all bones and produces localized agony, is extraspinal tuberculous osteomyelitis. While neighboring tissues are affected, problems such as paralysis of the face, tenosynovitis, and syndrome of the carpal tunnel may result [47]. A chest MRI shows lung disease in 50% of those with osteoarticular TB, although severe respiratory infection is rare [25]. The degree of decay of bones, softer tissue growth, and invasion on surrounding structures like the vertebral column may all be assessed by an MRI test [45, 46]. Making a determination of skeleton or articular TB requires a high level of probability.

When people have a passive medical history that includes osteomyelitis affecting the spine of the chest or monoarticular infection with negative microbiological samples, specialists should check them for spinal TB. Up to 80% of patients with tuberculous arthritis had positive results from arthrocentesis with synovial fluid mycobacterial culture. A synovial biopsy may also be used to diagnose a disease (caseating granulomas on histology or a positive mycobacterial culture) [26,27]. For the diagnosis of tuberculous osteomyelitis, a skeleton sample must be obtained for histology and bacterial cultures, and surgery may be required to get rid of infections, remove infected cells, stabilise the vertebrae, and relieve nerve tension [28]. In the lack of neurological damage, a weakened vertebra, or nerve compression as well, treatment with medicine alone (see Principles of Management)8,28 ought to yield a positive outcome.

Pathophysiology: Mycobacteria thrive in environments with plenty of oxygen. Therefore, they infect the spine when they spread through hematogenous means to the central and anterior regions of the vertebral bodies. The infection results in spinal degeneration and intraosseous cold abscesses. Due to the body's insufficient and gradual inflammatory response, the bacilli move to nearby or distant vertebral bodies through the Batson venous plexus. Mycobacteria lack proteolytic enzymes, resulting in the preservation of the intervertebral disc [39,40].

4. Central Nervous System Tuberculosis: Instances of TB in the brain and spine include spinal tuberculous arachnoiditis, cerebral tuberculomas, and tubercular meningitis (the most common form). A subependymal hump breaks into the cerebrospinal space, which results in severe irritation and causes meninges. [29] The arachnoiditis that ensues protects the entering arteries and cranial nerves, causing communicative encephalopathy and spinal palsies.

Vasculitis of the skull can cause localized neurologic impairments. Meningismus and characteristic cerebrospinal fluid (CSF) findings can be caused by tuberculo-protein hypersensitivity. While tuberculomas might present as space-occupying lesions, cerebral edoema causes dizziness, seizures,

and elevated intracranial pressure [30].

Chronic headaches, meningitis and vomiting, sense of disorientation and localized neurological irregularities appear approximately two weeks after fatigue, a high fever, or behavioral alterations. If untreated, mental illness might progress to lethargy or unconsciousness [31]. A quick progression that resembles slides meningitis or modest cognitive decline that resembles dementia, and a predominant illness that resembles encephalopathy are examples of unusual characteristics. Convulsions are might occur at any point when the condition is present. A strong degree of suspicion is necessary for quick evaluation and therapy start-up. The CSF has a mild lymphocyte pleocytosis (one hundred to five hundred cells per RL [0.10 to 0.50 10⁹ cells per L] [16], originally predominating in neutrophils).

CSF proteins can be high enough (2 to 6 g per dL [20 to 60 g per L]) in patients with xanthochromia having cerebrospinal block, ranging from 100 to 500 mg per dL (1,000 to 5,000 mg per L). Typically, the CSF has a sugar content of below 45 mg/dL (2.50 mmol/L). AFB smears on CSF are positive in 10–90% of patients, although susceptibility can be increased by analysing significant volumes of fluid from many spinal injuries, centrifuging it into CSF, and performing an AFB smear on the pellicle, or by having a skilled investigator look at a number of strong-powered fields. A 45 to ninety percent people have an elevated infection with M. tuberculosis CSF culture, although it typically takes a period of four to six weeks.

[32] In the proper clinical situation, an increased CSF adenosine deaminase level supports the diagnosis. [33] On a CT scan of the head with contrast, basilar arachnoiditis, infarction, hydrocephalus, or tuberculomas may be identified. When clinical, laboratory, or imaging signs imply tuberculous meningitis, empiric anti-tuberculous therapy should be started. Delay in starting therapy has been linked to poor outcomes. For at least 9 to 12 months, anti-tuberculous medication should be continued. [8] Adjunctive corticosteroid therapy with dexamethasone (Decadron) over the first six to eight weeks has been associated with reduced mortality and fewer neurologic sequelae in individuals with tuberculous meningitis [8,11,34] Patients under the age of five, those over the age of 50, and those who have been sick for more than two months have the highest mortality rates.

Pathophysiology- Basal meningeal elevation, hydrocephalus, and parenchymal infarction are among the imaging signs of tuberculous leptomeningitis. These results are very specific for tuberculous meningitis [11,12]. In the basal cisterns, viscous exudates with a high protein concentration are common [11]. Hydrocephalus is caused by inflammatory exudates that restrict CSF flow in the subarachnoid space. The condition is viewed as a side effect of tuberculous meningitis [11]. About 20% of people with tuberculous meningitis experience localised neurologic deficits as a result of brain infarctions. The most widely acknowledged pathophysiologic mechanism involves submerging the lenticulostriate and thalamoperforating arteries in an inflammatory meningeal exudate, which harms the adventitia and causes vasculitis [13]. Intimal proliferation with stenotic disease and fibrinoid vessel wall necroses have also been seen in patients with TB meningitis and stroke [14].

5.Gastrointestinal Tuberculosis: Tuberculous enteritis can be brought on by inhaling contaminated sputum, ingesting tainted food, hematogenous spread, and direct extension from neighbouring organs[35]. The three most common forms of intestinal lesions are ulcerative, hypertrophic, and ulcero-hypertrophic lesions. Symptoms include fever, diarrhoea, weight loss, and abdominal pain. Symptoms

can include melana, rectal bleeding, and stomach discomfort. A mass in the right lower quadrant might be felt in 25 to 50% of patients. The ileocecal region and the jejunioileum are the most often involved areas. Complications include obstruction, perforation, and the development of a fistula. The most frequent rectal lesions are perirectal abscesses, anal fissures, and fistulas [16]. During barium contrast tests and colonoscopies, it is possible to find fistulas, strictures, a deformed cecum, and ulcers.

An abdominal CT scan can find extraluminal pathology, especially lymphadenopathy. It is crucial to distinguish tuberculous enteritis from Crohn's disease since the start of immunosuppressive therapy in a patient with tuberculous enteritis could promote dissemination. A definite diagnosis is made using histopathology and culture of biopsy samples obtained by colonoscopy or laparotomy. The differential diagnosis for tuberculous enteritis include Crohn's disease, amebiasis, neoplasia, Yersinia infection, and actinomycosis [35]. Six months of antituberculous therapy are indicated. Only those with difficulties are candidates for surgery.

Pathophysiology- After being coughed or ingesting tainted milk, the bacilli infect the gastrointestinal tract. Hematogenous spread is also a possibility. The bacteria attack the lymphoid tissue beneath the mucosa, causing submucosal granulomas that can subsequently ulcerate. The surrounding mucosa thickens due to the inflammatory response. The terminal ileum is more frequently impacted due to a variety of contributing variables, including intestinal fluid stasis, the presence of significant lymphoid tissue, higher absorption rates, and closer contact between the bacilli and the mucosa [32]. TB, on the other hand, can impact other GI tract regions. On a CT scan, necrotic lymph nodes as well as asymmetric wall thickening of the terminal ileum, cecum, or ileocecal valves may be visible. The cecum may appear small and irregular following persistent inflammation due to fibrosis and stenosis [32].

6.Tuberculous Peritonitis:Persons who have a liver disease or infection with HIV and are receiving ongoing peritoneal dialysis treatment are more likely to develop tuberculous peritonitis, 35 Tuberculous peritonitis is brought on by the stimulation of persistent sites in the peritoneal. Fever, which is fluid retention, and abdominal pain are all signs that develop progressively. Peritoneal fluid is exudative (11 g per L) when the serum-ascites albumin gradient is smaller than 1.1 g per dL. A lymphocytic predominance is observed [16] when tuberculous peritonitis complicates continuously ambulatory dialysis in the peritoneum; a neutrophilic pleocytosis is observed [16] when tuberculous peritonitis complicates continuously ambulatory dialysis in the peritoneum [36].

By centrifuging 1 litres of peritoneal fluid, it is possible to increase the yield of AFB smear and mycobacterial culture. The adenosine deaminase level in peritoneal fluid has good sensitivity and specificity if a threshold value of greater than 33 U per L (550 nkat per L) is used. A peritoneal biopsy aided by laparoscopy or a mini-laparotomy can be diagnostic in more than 95% of patients, thus it should be carefully examined.

Solid-Organ TB Abdominal: The two organs that are most frequently impacted by solid-organ TB are the liver and spleen. Immunocompromised individuals are most frequently affected, particularly those with severe HIV infection [35]. Pathophysiology— In the liver and spleen, micronodular or macronodular symptoms can be seen. They are frequently associated with miliary pulmonary

tuberculosis and spread to the belly by hematogenous dissemination [35].

7. Genitourinary Tuberculosis: Direct infections of the kidneys or lower urinary tract or subsequent amyloidosis are both potential causes of renal illness. Dysuria, hematuria, and flank pain are common symptoms. [37,38] A sterile pyuria with or without microscopic hematuria affects more than 90% of asymptomatic patients. [37,38] A "moth-eaten calyx" or papillary necrosis can be seen on intravenous pyelography [99]. An abdominal computed tomography (CT) scan may identify pulmonary calcium deposits, calculi, damage, water retention, or extra renal disease-related symptoms such urethral obstruction, bladder constriction, or calcium deposits in the prostatic, seminal, or venous systems.

A mycobacterial culture of three morning urine samples confirms the diagnosis in 90% of cases [21,37]. For prolonged flank pain or hypertension, nephrectomy is rarely recommended. [8] Except in the case of tuberculous interstitial nephritis, renal function is normally retained [22]. Male genital tuberculosis frequently coexists with renal TB. It first affects the prostate before moving on to the seminal vesicles, epididymis, and testes. Patients frequently have a scrotal mass when they first present, and surgery is used to identify the problem. Oligospermia is a common disorder that can linger for a very long time. Beginning in the endo salpinx, female pubic TB can progress to the peritoneum the endometrial reproductive organs, the cervical cavity, and the vaginal area. [38] Usual signs include bleeding out of the vagina, being infertile, and abdominal discomfort.

All kinds of genital TB respond well to chemotherapy, however severe tubo-ovarian abscesses in women necessitate surgery [8].

Male Genital TB: Genital TB in men is rare and typically accompanied by renal tuberculosis. The most often impacted organs are the prostate and epididymis. Epididymitis and isolated testicular TB are frequently associated [26,27].

Pathophysiology- Hematogenous spread is the male genital TB mechanism that occurs most frequently. Although it occurs more commonly, prostate tuberculosis frequently has no symptoms. Most cases of prostate TB are found by accident when a prostate biopsy is performed to rule out malignancy. Prostate specific antigen levels can dramatically rise, and urine analysis reveals sterile pyuria [27]. Renal TB and retro canalicular hematogenous spread from prostatic illnesses are both potential causes of epididymal tuberculosis [28].

Female Genital TB- Despite its rare, vaginal TB in females is a pathophysiological factor in sterility. Typically, the endometrial is compromised first, then the tubes of fallopian[29].

Up to 25% of infertile women may have genital tuberculosis in areas where the disease is prevalent [29,30]. It is impossible to exaggerate the value of diagnosis. When given anti-tuberculous drugs and assisted reproductive techniques, more than half of genital TB patients' women may be able to conceive [30].

8.Miliary Tuberculosis:A dispersed, chronic type of TB known as miliary tuberculosis may occur during the first spread or after decades of uncontrolled disease. 10% of patients with the HIV virus with

pulmonary TB and 38 percent of people having AIDS and extrapulmonary TB had mild disease. [5] The most common signs include eating disorders, losing weight, night sweats, a high temperature, and shivering. The signs of illness depend on the implicated organs. Fulminant illnesses include multiple organ failure, intense breathing difficulties condition, and bacterial infection.

In more than 85% of patients, A computed tomography (CT) scan or chest MRI reveals several 2- to 3-mm lesions scattered across the lung's wall. [16,39] Typical testing findings include normochromic anaemia, leukaemia or anaemia, elevated deposition rate, and dehydration. For an identification, 40 it may be necessary to do a throat inspection, bronchoalveolar lavage, gastrointestinal washings, bloodstream fluid, cultured blood, or hepatic and blood vessel biopsy[16]. A tuberculin skin test is only positive in roughly 50% of tuberculosis patients. As previously indicated, anti-tuberculous drugs are used to treat tuberculosis, as well as corticosteroids in some situations.

Pathophysiology-Baseline meningeal the elevation, hydrocephalus that and parenchymal inflammation are among the diagnostic signs of tuberculous leptomeningitis. The outcomes are extremely specific for tuberculous meningitis. [11,12]. In the fundamental cisterns, discharges with a considerable stickiness and high level of protein are common [11]. Hydrocephalus is caused by inflammation fluids that restrict blood circulation in the cerebral region. As a result of tuberculous meningitis, it is thought to have repercussions [11]. As a result of cerebral infarctions, about 20% of those with the condition have localised neurological problems. The most frequently acknowledged pathophysiologic process involves an inflammatory meningeal discharge submerging the lenticulostriate and thalamic-perforating veins, which damages the portal vein and results in vasculitis [13]. Additionally, those with TB meningitis and infarction had intimal development with stenotic disease and fibrinoid wall deposits [14].

9.Tuberculous Pericarditis: Continuous transmission of tuberculosis from the mediastinal nodes, lungs, spine, or sternum, as well as miliary dissemination, are the two main causes of tuberculous pericarditis. Chest discomfort, dyspnea, and ankle edoema are examples of symptoms that might arise suddenly or gradually. A cardiomegaly, tachycardia, fever, pericardial rub, pulsus paradoxus, or enlarged neck veins may be discovered during an examination. Compared to pericardial fluid alone, pericardial biopsies are more likely to result in a precise diagnosis[41]. Anti-tuberculous medication and corticosteroids are both recommended to hasten symptom resolution and reduce fluid re-accumulation [8,41]. Constrictive pericarditis risk or mortality are unaffected by corticosteroids. [11] Open pericardial drainage is preferable to pericardiocentesis. [41]

Pathophysiology-In the liver and spleen, micronodular or macronodular symptoms might be seen. They are frequently associated with miliary pulmonary tuberculosis and spread to the belly by hematogenous dissemination [35].

10.Tuberculous Osteomyelitis:appears an instance of TB that is less prevalent than tuberculous arthritic. While multicenter osteomyelitis is less prevalent in youngsters, unifocal invasion can nevertheless happen. The thighbone, the tibia, and little bones in the palms of your hands and toes are often impacted joints. [34].

Pathophysiology -Hematogenous spread allows the bacilli to enter the long bone metaphysis. Granulomatous lesions develop into necrotic caseating nodules. Trabecular resorption is also a possibility. The joint space and epiphysis could both get infected as a result of the infection. Additionally, it may result in the cortical bone disintegrating and spreading to neighbouring joints or muscles [44].

TNF-G INHIBITOR-ASSOCIATED TUBERCULOSIS: Infliximab (Remicade) [42] and etanercept (Enbrel) [43], both TNF-G inhibitors, have been linked to active tuberculosis in patients with rheumatoid arthritis or Crohn's disease, according to two recent reports. It took less time to identify tuberculosis after starting infliximab than it did after starting etanercept (median of 12 weeks versus 12 months). According to [42,43](Fig. 2), extrapulmonary tuberculosis was the cause of 52 to 57% of cases. Therefore, before beginning TNF-G inhibitor therapy, patients should be evaluated for latent tuberculosis infection or current tuberculosis disease [98].

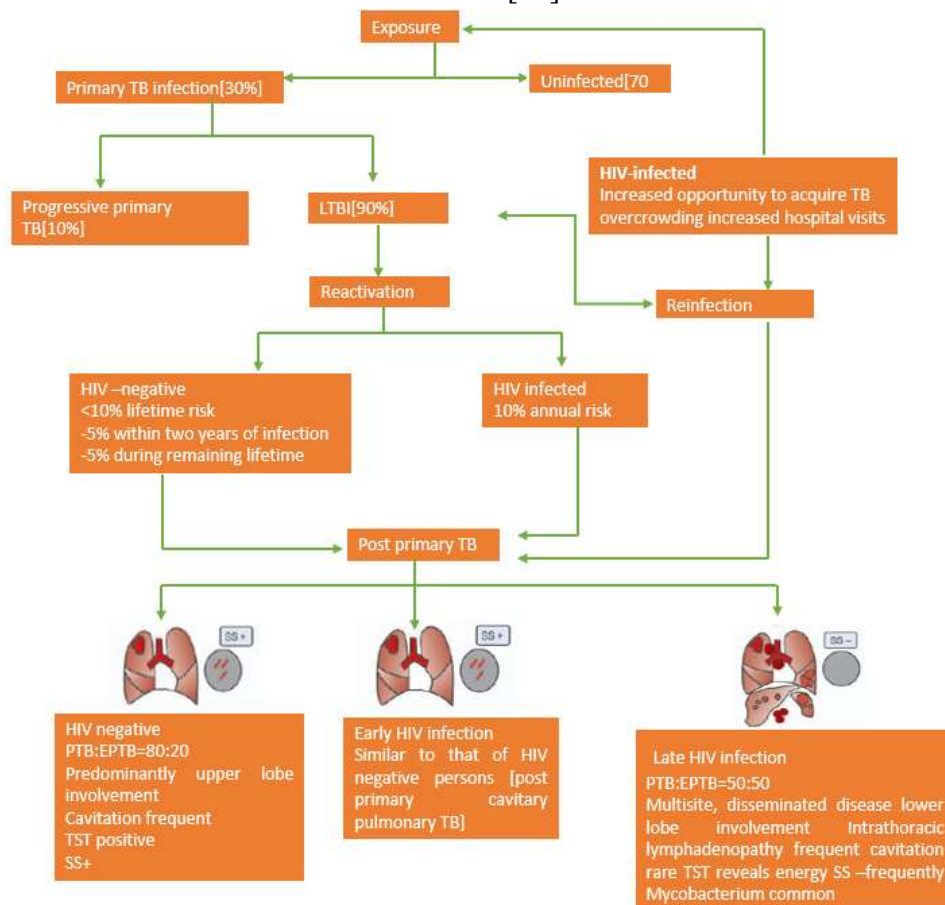


Figure 2: Overview of the Extra Pulmonary tuberculosis infection

Diagnostic Tools for Extra Pulmonary Tuberculosis:

Extrapulmonary tuberculosis (EPTB) accounts for about 20% of all tuberculosis (TB) cases in India. Diagnosing EPTB is still challenging because clinical samples obtained from particularly hard-to-access

locations may be paucibacillary, reducing the sensitivity of diagnostic testing. When possible, every effort should be made to obtain appropriate samples for both mycobacteriologic and histopathologic studies. Measurements of adenosine deaminase, gamma interferon, and molecular biology techniques like polymerase chain reaction in TB-affected serosal fluids may be useful adjuncts in the diagnosis of EPTB. Despite the fact that the illness frequently responds to standard anti-TB pharmaceutical therapy, the best regimen and duration of treatment have not yet been found. Treatment for anti-TB frequently causes a paradoxical effect. It must be distinguished from further clinical worsening factors. The major reasons for needing surgery are to collect reliable diagnostic samples and to treat problems. Clinical monitoring is typically used to gauge a patient's response to treatment since smear microscopy or culture cannot be used to monitor EPTB patients [108].

NAA tests: The development of NAA tests, including polymerase chain reaction (PCR) that identify genomic sections specific to MTB right away in extrapulmonary specimens, which provide outcomes within a matter of hours, provide more accuracy than AFB microscopy of smears, and more frequently than the culture of Bacteria [109,27], resulting in an important development in the identification of EPTB, particularly for medical circumstances with a greater risk of HIV-EPTB combination [109, 112, 27, 49, 96, 97]. The main objective of the current study is to create PCR as a diagnostic tool for a variety of clinical variations of EPTB using various criteria. The 65kDa peptide (Rv0934), TRC4 (conserved repeating motif), GCRS (guanine-cytosine-rich repetitive motif), hupB (Rv2986c), dnaJ (Rv0352), MTP -40 protein (Rv2351c), and PPE gene (Rv0355) are the targets of these polymerase chain reaction assays. [120, 58, 90, 91, 96, 97]. More than one copy of the complicated chromosome for mycobacteria are present, which can increase susceptibility. In tests using PCR, IS6110 was often utilised [115, 144, 104].

certain investigations across various geographical locations indicate that certain clinical samples have either just one duplicate of IS6110 or none at all, leading to erroneous negative indicates [79,166]. Nested PCR and multiplex PCR were used to increase efficiency and prevent interference while replication (two-step replication [167, 116, 51] and parallel replication of more than one genes target [134, 76, 155, 156] have been utilised to diagnose EPTB). A positive mRNA signal suggests the existence of bacteria since the DNA-PCR cannot differentiate among functional and infertile pathogens and since bacteria mRNA have a shorter duration of action than genetic DNA [145]. Use mRNA-based reverse transcriptase-PCR (RT-PCR) to quickly distinguish between *M. tuberculosis* strains that are alive and those that are not. It is also used to detect EPTB and keep track of treatment resistance [85,145]. Real-time PCR is a novel and trustworthy technique for measuring the DNA components in EPTB samples. The primary function of real-time PCR, an innovative and reliable method, is for measuring the DNA components in EPTB samples [58, 148].

The primary benefits of employing this actual time PCR test include a shorter reaction time, measurement of microbe load, automating of the process that cuts down on practical effort, and a lower danger of getting contaminated by other bacteria [105,148]. Numerous inhibitors, such as host proteins, blood, and even eukaryotic genetic material in extrapulmonary materials, interact with PCR

responsiveness, which is the main reason why PCR amplification results are frequently erroneous [89,96,97,61,62]. In most cases, an extensive procedure is needed to get rid of inhibitions of PCR and yield very pure Material. A variety of techniques for processing DNA specimens have been proposed to achieve this, including thaw-boiling, chelex/proteinase K therapy, and The technique of pattern capture. Chakravorty and Tyagi (2005) [69] proposed a novel adaptable universal sample preparation (USP) method that could be used for blocker-free PCR in both PTB and EPTB samples, using the chaotropic feature of guanidinium hydrochloride as a major component. Additionally, it is being shown that introducing silicon films or cetyltrimethylammonium bromide to the purging of DNA procedure efficiently eliminates inhibitions of PCR, possibly improving PCR sensitivities in EPTB samples[60, 101, 144].

A brief cultured amplification procedure lasted two or three days is being advised prior to the PCR analysis to avoid this issue since the extra purification steps might cause a large loss of mycobacterial DNA[60,101,144]. This might increase the mycobacterial burden while concurrently reducing the effectiveness of PCR inhibitors, as suggested by Cheng et al., 2005 [72, 73]. In a clinical case of pleural tuberculosis, nine distinct DNA extraction techniques (seven manual and two automated) were put to the test by Santos et al. Only a few methods—column segregation (Qiagen) and automated extracting with complete separate nucleic acid (Roche) system—were successful in detecting the M. tuberculosis Gene in each of the pulmonary TB specimens, suggesting that the method used to collect genetic material may have an effect on the efficiency of real-time PCR. Due to the PCR's high sensitivity, residual contamination of the amplicon, prior infection, or ongoing EPTB illness at another place could result in a false-positive result [101,69, 70,61, 62]. False positive PCR results in the absence of clinical symptoms are posing severe issues in diagnosing EPTB infections these days [166].

Evaluation of innovative diagnoses in EPTB individuals is still significantly hampered by the absence of a good standard of reference [61, 62, 171]. The actual precision of PCR analyses may vary from what is stated if a subpar gold/reference benchmark is used [49,168]. Cultured (on solid and liquid media) is the most frequently employed gold standard for confirming PCR results for identifying EPTB samples, despite the fact that it is a substandard gold standard with variable susceptibility and leads to incorrect PCR results [127, 128, 100, 61, 62]. The additional gold/reference criterion includes BACTEC the culture, histology, and outcome of anti-tubercular treatment (ATT), as well as an assortment of these assays [127, 128,109,50,130]. Chakravorty et al. (2005) used smear, culture, pathogenesis/cytology, clinical signs, and responses to ATT [69] and Vadwai et al. (2011) [171] as the gold/reference standard for verifying the PCR outcomes for identifying EPTB samples.

Tuberculous lymphadenitis: Approximately 35 percent of EPTB patients exhibit this tuberculous lymphadenitis, making it among the most prevalent EPTB diagnosis [124, 77, 80]. The collar lymph glands have become the first to be affected by this medical condition, proceeded by the mediastinal, axillary, mesentery, liver opening, and genitals lymphatic cells [151]. Since lymphadenitis from TB resembles various medical conditions (such as fungal infections, sarcoidosis, , NTM infections and leprosy) and shows variable histology results despite the lack of AFB, TB lymphadenitis can be difficult

to diagnose [81]. In order to diagnose TB lymphadenitis, FNA cell biology, a less invasive procedure than elimination biopsies, is growing and increased its significance [69, 70, 81]. To carry out an AFB smear and culture study, however, typically requires more information than is retrieved after the FNA [124]. Additionally, TB can be challenging to differentiate with other granular and or NTM diseases using FNA cytology [55]. Several genome groups, particularly, 16S rRNA gene, 65 kDa, IS6110, MPB-64 and IS1081 are being utilised to identify TB lymphadenitis by Amplification using FNA or formalin-fixed paraffin-sub-merged samples with varying susceptibility and particular characteristics (169, 137, 135, 129, 155, 156; 77; 81; Table 1). The medical use of amplification by PCR combined along with FNA cytologists may lessen the requirement for readily accessible biopsies, since it's incurable and causes unsightly wounds in the neck area that cause visual problems [55, 163]. A number of investigators utilised the remainders of FNA immediately following cytological testing to conduct PCR. One of the difficult TB problems *M. bovis* and *M. tuberculosis* are both major causes of TB lymphadenitis [154].

In order to detect Mycobacteria at the genera (Antigen 85 complex gene), complex (IS6110) species (*pncA* gene and allelic variation) levels in TB lymphadenitis patients, the leftover FNA has been employed for PCR based on three genome domains during cell biology study. Eighty-seven percentage of the samples found were confirmed by PCR positive for *M. TB* in the genera and complicated level, a percentage of 68.5 at the scale of species, and seventeen percent for *M. bovis*. A networked PCR addressing the *hupB* gene was previously shown to distinguish MTB with *M. bovis* with FNA samples [174]. A PCR analysis relying on person *devR* and targets IS6110 indicated an average susceptibility in USP-processed lymphatic node specimens compared to patients with TB lymphadenitis, but the level of specificity could be improved through merging the outcomes of the two tests (by running different PCR reactions for every single gene target; 69, 70). Employing an intersection of 65 kDa PCR and IS6110 data, Mirza et al. (2003) [123] studied the identification of TB lymphadenitis in mononuclear cells in peripheral that showed greater susceptibility to lymphatic node in Amplification. NTM lymph node infection seems to be a novel disease in children. A rapid PCR technique was created to facilitate a fast determination of this illness relies on the internal transcribed spacers a series (between the 23S rRNA gene the 16S rRNA and), facilitating the recognition of the genera Mycobacterium and both species Mycobacterium tuberculosis and Mycobacterium avium [63]. The good outcomes of their approach for finding unusual mycobacterium should be helpful for individuals with lymph node infection deciding medical choices.

Pleural tuberculosis: 3-25% of people with tuberculosis have pulmonary TB, with TB the condition representing the greatest frequent reason for pleural drainage [114, 116]. It is inefficient and laborious to diagnose pleural TB by looking for bacilli with tubercles in fluid from the pleura and biopsies samples, or by finding granules in pleural region. The insufficient yield of microscopy (culture and the intrusiveness of pleural biopsies have renewed interest in potential noninvasive treatments [114]. Pleural drainage can result in elevated concentrations of interferon- γ (IFN- γ) and adenosine deaminase (ADA) which can be utilized to identify pulmonary TB [175, 105]. With an accuracy of > 91%, Sharma & Banga (2005) [150] proved the tests' usefulness in TB pleural effusions. Due to the costly nature of

IFN- γ test, ADA test is preferred than IFN- γ test in countries with limited resources; yet, ADA analysis is being reported to be good for various illnesses like lymphomas, collagen vascular disorders and adenocarcinomas, [113,115].

Utilising genetic markers with varying ranges and specificities, including MPB-64, GCRS, *devR* and IS6110 the effectiveness of PCR for the identification of TB pleural effusions is being widely studied [120, 69, 70, 96, 97]. Incorporating the findings of the *devR* and IS6110 PCR assays, Chakravorty et al. (2005) [69] reported a high specificity in fluid from the pleura and cannula-biopsied pleural bone. A novel repeating repetitive region called CD192 was recently found in the *M. tuberculosis* DNA, and PCR has effectively utilized it to distinguish between pleural TB and TB meningitis [159]. Over 300+ tuberculosis clinical individuals as well as those lacking IS6110 patterns, contained CD192 motifs. Many investigations have looked at multiplexed or live PCR assays to boost accuracy. Applying multiplex PCR techniques directed against the IS6110, *dnaJ*, and 65 kDa protein genes, tuberculosis was found in the peritoneal fluid, pleural fluid and CSF [58]. Greater specificity could result from the application of monoplex/multiplex PCR which works in combination with ADA estimations or histopathologic analysis of pleural specimens [115,116,58]. For the detection of pleural TB in formalin-fixed paraffin-embedded pleural tissue, a quick PCR approach addressing the 65 kDa peptide gene was created, and the test's specificity was equivalent to a nested PCR targeting IS6110 [54]. On the contrary hand, Rosso et al. (2011), [148] gained little specificity with rapid PCR in pulmonary TB patients, although their findings that the results remained better than AFB screening and culture. Villegas et al. (2000) had before established a high degree of specificity and sensitivity for the rapid identification of pulmonary TB with regard to positive PCR or ADA/IFN- γ findings. Similar results were presented just recently by Kalantri et al. (2011) [105], who asserted significant accuracy (96–100%) in determining the presence of pulmonary TB via significant rapid PCR or IFN- γ findings

Tuberculous meningitis: The most prevalent and deadly kind of meningitis in developing countries is tuberculous meningitis, which affects 7–12% of those with TB [110]. Postponing therapy may cause permanent harm to the brain and TB meningitis carries a nearly 100% fatality rate [165, 153, 155, 156]. As a consequence, for successful treatment care, timely identification of TB meningitis is essential. PCR is now routinely utilized to identify *M. tuberculosis* in CSF utilizing IS6110, 65 kDa, 38 kDa, *devR*, MPB-64, or PPE gene targets and varied specificity while traditional diagnostic procedures for TB meningitis nearly never succeed [120, 110, 143, 159, 144, 83,165, 96, 97]. Furthermore, *M. tuberculosis* Genome in CSF is detected by PCR, which has a better accuracy than computational tomography (CT) scanning since CT scanning only identifies. The comparative effectiveness of all three PCR tests—IS6110 PCR, nested PCR, and MPB-64 and 65 kDa peptide genetic targets—was evaluated in a single CSF the specimen, 144. Other IS6110 PCR, a single-step tests, has the advantage of offering a fast test for the identification of meningitis caused by TB with a higher degree of specificity and sensitivity in contrast to the nest techniques. The three different genes targets (IS6110, MPB-64, and 38 kDa protein genes) were subsequently combined into an effective multiplex PCR technique by Sharma et al. (2011a) [155, 156], and the results surpassed the single plex PCR using only one genome targeting.

Obtaining an adequate amount of CSF collection from paediatric patients is challenging, affecting the identification of TB meningitis complicated [110, 88]. The amount of the CSF specimen is essential for reliable PCR findings. A effective PCR method for the diagnosis of TB meningitis among kids was disclosed by Kulkarni et al. (2005) [109] using a little amount of the total CSF and focusing on the 38 kDa peptide genes. Their method had the ability to identify 10 femtograms (fg), roughly 2-3 tuberculosis bacilli, of Genetic. Rafi et al. (2007) [144] used the 'full' CSF rather of the sediments in the PCR investigate, showing that *M. tuberculosis* Genetic may be detected in the CSF specimens as independent DNA fragments. Haldar et al. [96, 97] demonstrated the effectiveness of CSF' filtration for identifying *M. tuberculosis* Genome employing both immediate PCR and traditional PCR addressing the IS6110 and *devR* genes. unexpectedly both analyses showed that CSF 'filtrate' exhibited better specificity as well as sensitivity over sedimentation. Their approach proved especially useful for assessing the medical records of individuals having meningitis caused by TB on ATT. They used the method of quantitative nest RT-PCR (QNRT-PCR) technology to identify *M. tuberculosis* Gene in CSF specimens. A unique, thoroughly designed quantitatively nested RT-PCR (WR-QNRT-PCR) assay targeting the MPB-64 peptide genes is being developed to identify *M. tuberculosis* genome in CSF samples with a large detection range (1-105 copy counts) throughout the clinical development of illness [157, 165].

Osteoarticular tuberculosis: One to three percent of the instances of mycobacteria are osteoarticular, and it is the primary manifestation of osteomyelitis [179, 61, 62]. Each bones, socket, or tissue may become wounded, although the vertebral column, hip regions and elbow are particularly often affected, accounting for between 70 and 80% of illnesses. Spinal tuberculosis this, if not properly diagnosed and cleaned up, may result in kyphosis and/or neurologic problems that cause paraplegia [102]. Accurate diagnosis of osteoarticular TB is difficult because of wide, unreachable symptoms and the widespread initiation by experimental ATT [172]. Particularly in endemic locations, osteoarticular TB is often identified according to medical belief and imaging proof [51, 61, 62]. PCR analyses relying on the 16S rRNA gene, IS6110, targets and 65 kDa protein gene, were employed often for verifying osteoarticular TB, varying in specificity [173, 128; 102, 51, 61, 62].

Due to extensive introduction of experimental ATT and wide-spread, unreachable infections, accurate diagnosis of osteoarticular TB is challenging [172]. Osteoarticular TB is frequently determined to be present based on clinical symptoms and imaging evidence, especially in endemic areas [51, 61, 62]. PCR tests according to IS6110, 16S rRNA genes, and 65 kDa peptide genetic sequences are being utilised often, with a range of sensibility, for verifying osteoarticular TB [173, 128, 102, 51, 61, 62]. While Chawla et al. (2009) [71] suggest maintaining those specimens in regular water instead of formalin because that substance has been linked to DNA modifications, performing PCR on formalin-fixed, paraffin embedded cells is currently recorded in identifying evidence of tuberculosis osteomyelitis [103].

Additionally, it was shown that osteoarticular TB may be diagnosed using embedded PCR addressing IS6110 and mycobacterial culture, including automatic and traditional (51). A very accurate and precise multiplex PCR identifying the IS6110 and MPB-64 peptide sequences was used by Sharma et al.

(2011b) [155, 156] to analyse synovial fluid (SF) and abscess specimens from eighty individuals with osteoarticular TB [179]. Researchers created the *rpoB* PCR-plasmid TA replicating-sequencing technique to detect *M. tuberculosis* in joint cells, synovial fluid, and abscess tissues from osteoarticular TB. The method they created might also be used to determine how susceptible tubercle bacilli are to the antibiotic rifampin (RIF). Fujimoto et al. (2010) [87] identified an instance of TB pleura with invasion of the joint in the knee using PCR examination of synovial fluid. An enclosed region of the genome that codes for a single immunologic cellular protein of 31 kDa of *Brucella abortus* (BCSP31) and SenX3-RegX3 (intergenic region of *M. tuberculosis*) was used by Colmenero et al. (2010) [76,139] to perform a quick detection of TB vertebral osteomyelitis and brucellar vertebral osteomyelitis.

Genitourinary tuberculosis: Vaginal and kidney tuberculosis (TB): Representing as much as 46 percent of the overall incidence of EPTB, urogenital TB is the next most common kind [24, 27, 28]. Up to twenty times the number of people who receive kidney transplants than people, in general, are affected by renal infection. To prevent damage to the kidneys, hepatic tb must be detected earlier [176]. Sun et al. (2010) [161] suggested an immediate PCR method for detecting renal TB from kidney biopsy samples using 35- and 40-cycle threshold (CT) cut off values. The real-time PCR (CT 40) was shown to have a higher level of specificity than the real-time PCR (CT 40). (CT 35). Since endometriosis is silent and is frequently associated to vaginal TB in both men and women, a lot of instances are untreated. For a consequence, the identification of urogenital TB requires an elevated level of suspect. PCR targeting the MPT-64 peptide genes was already shown to be the most accurate method for detecting gastro intestinal TB in both men and women) in specimens of urine, superior to intravascular urography, urinary biopsies, or urine cultures. The effectiveness of PCR addressing the IS6110 or 16S rRNA genome for the identification of gastro intestinal TB in specimens from urine has additionally been studied [126, 49].

Relying on the MTP-40 peptide genes of *Mycobacteria* TB, nesting PCR has been reported to have as much as one hundred percent accuracy. [90]. A number of specimens were employed by Bhanu et al. (2005) [59] to demonstrate the screening capability of PCR addressing the MPT-64 peptide genome in detecting the tubercular cause of female impotence. These samples included endometrium aspirates, endometrium biopsy specimens, and secretions from the pouches of Doug. Additionally, they compared the surgical outcomes with the PCR data. According to Thangappah et al. (2011) [166], TRC4-based PCR had more specificity (91-100%) than IS6110-based PCR for identifying clinically probable instances of female gastro intestinal TB in test specimens. It also has a higher degree of specificity than IS6110-based PCR. For a quick identification of PTB, Amplification was recently utilised to identify mycobacterial transrenal Gene in specimens of urine [167, 93].

Abdominal tuberculosis: Abdomen tuberculosis: The global epidemic of HIV has significantly increased incidence of abdomen TB, accounting for 10-12% of EPTB patients [66]. Thoracic TB is defined as TB of the gut, peritoneum, mesentery, and other integration-abdominal organs like the liver, spleen, and pancreatic [151]. Because histological poses an identification challenge, the use of DNA for the detection of abdomen TB has been used, and PCR may additionally assist in ruling out carcinoma

in fresh surgical abdomen specimens [111]. Kulkarni et al. (2006) [109] reported that abdominal TB could be detected by PCR with considerable specificity and sensitivity utilising the 38 kDa peptide genes, and the PCR diagnostic was additionally incorporated with an Indian professional package [111,160]. They used histological as the standard of excellence [89].

In TB-endemic nations where Crohn's disease is on the rise against a backdrop of higher intestinal TB occurrences, it is imperative to be able to differentiate between these two diseases [52, 84, 140]. Gan et al. (2002) [89] discovered that PCR is a good diagnostic in the distinction of intestinal TB and Crohn's disease, but biopsies has low accuracy in the distinguishing of the two disorders. An internal PCR (targeting IS6110) from endoscopic biopsy specimens was compared with two commonly used commercialised PCR packages, kit (targeting MPB-64 and IS6110) and kit (targeting IS6110), for the diagnosis process of these two disorders [104, 142]. It was discovered that gastrointestinal TB and Crohn's illness may be distinguished from one another using PCR using a kit and histopathologic analysis. Balamurugan has discussed how to discriminate between both of these diseases using a fecal coliform bacteria-TB PCR test that targets IS6110.

Yet, the clinical application of this PCR test is yet to be demonstrated in a sizable patient population. Conventional PCR has a number of drawbacks, one of which is the requirement for tissue damage and DNA extraction, which prevents comparison to histological features [162]. An in situ PCR that coupled the ability to detect specific DNA across tissues replicated the IS6110 target throughout whole cells. This technique might also separate gastrointestinal TB with Crohn's disease in historical membrane biopsies samples. Once its usefulness is confirmed, in situ, PCR's sensitivities must be improved, and research on a substantial percentage of people with gastrointestinal TB and Crohn's disease has to be done [52].

Cutaneous tuberculosis: Skin tuberculosis: almost one quarter of EPTB infections are due to this condition. Still, during the past 20 years, PTB and multiple-drug-resistant tuberculosis (MDR-TB; 50) have become more prevalent, and this illness has returned [48]. The most common medical form is scrofuloderma, which follows by lichen scrofulosorum, psoriasis vulgaris, and TB verrucosa cutis. In contrast to the unfavourable outcomes of cultures and microscope tests, PCR was able to validate these clinical manifestations of corneal TB [136]. Strangely, Okazaki et al. (2005) [134] reported the first case of *M. bovis* BCG-derived skin TB (located away from the vaccination site) using a multiplex PCR test using the region of variance (RD) 1 supplement sequences RD1, RD2, RD8, RD14, and SenX3-RegX3 regions starting from *M. bovis* BCG. Tokyo 172 Amplification has found TB cuts orificialis as a rare sign of tuberculosis of the skin (produced by activate-inoculation of *M. tuberculosis* in individuals with very qualified and competent TB) [75]. With lifestyle/histopathology as the gold standard, it has been conclusively shown that IS6110-based conventional amplification/nested PCR is superior to 16S rRNA genotype-based PCR for the diagnosis of cutaneous TB [131,132]. A highly sensitive and exact PCR test targeting the 65 kDa peptide gene has also been developed for the detection of epidermal TB using culture/response to ATT as the gold standard [127,50].

Ocular tuberculosis (OTB): It is an uncommon variant of EPTB affecting 0.14–16 per cent of persons. It comes from localised damage and inflammation induced by haematogenous spread or an allergic reaction produced by distal infection with TB [53, 78]. The bulk of individuals with positive PCR findings have found medical recovery with ATT, and PCR has had confirmed to be an extremely successful methodology for the earlier recognition of ocular TB that it could be carried out with extremely tiny specimen quantities taken through the eyes [72, 73, 95]. A nested PCR targeting the MPB-64 peptide genes had previously been reported to be used in formalin-fixed paraffin-embedded material of the epiretinal membrane. The epiretinal membrane of an individual bacteria in Eales' illness could be identified with this technique, that identified 0.25 fg of DNA. The identical nested PCR test, however, was less sensitive in aqueous specimens [117, 118] (Table 1). The results of a recent study that examined the efficacy of real-time PCR methods using IS6110 or MPT-64 protein gene targets in the diagnosis of ocular TB are encouraging [155, 156, 178]. Additionally, M. tuberculosis was discovered in donated corneas using PCR assays. More research on the likelihood that recipients with PCR-positive corneas may ultimately result in infection transmission should be conducted since these findings may be utilized to reassess criteria for the acceptability of donations with active tuberculosis [67].

Pericardial tuberculosis: Pleural TB is the main cause of pericarditis in countries in Africa and Asia [74]. The causes of its development include lung tissue, military distribution, and spreading via mediastinal sites [92, 152]. Increased levels of both ADA and IFN- γ are being seen in cardiac TB [65], although as has been reported in pulmonary TB, these assays have drawbacks. Both traditional and nested PCR techniques have been recorded for identifying cases of acute pleuro pericardial TB and persistent constricting pericarditis [170, 180].

Thyroid tuberculosis: Thyroid tb: When there is a mega-nodular goitre, a mass, or a chronic sinus in the tissue, it is generally medically explored [64]. Chest X-rays and ultrasound are the two most often used tests for detecting basic thyroid TB, although they usually fall short. A multiplex PCR technique addressing the IS6110, 65 kDa, and dnaJ sequences was created to verify the presence of thyroid TB [91].

Tuberculosis mastitis: Even in regions where it is ubiquitous, chest TB, also known as tuberculous mastitis, is a rare indication of EPTB. A solitary, ambiguous, unidirectional solid lump in the breast's centre or top exterior region is the main indicator of breast TB [56]. Mycobacteria TB bacilli can reach the breasts by connected, haematogenous, or lymphatic spawning [151]. It was recently established that PCR may be used to diagnose chest tuberculosis [107].

Disseminated tuberculosis: The chronic and potentially deadly condition known as "disseminated tb" (sometimes called "miliary tuberculosis") is brought on by the broad hematogenous dispersion of tuberculosis causing bacilli in the human body [150, 88]. Spreading mycobacteria has been identified in 10% of AIDS-related PTB individuals as well as 38% of AIDS-related EPTB patients [92]. Spreading TB is challenging to diagnose clinically due to the fact that it can mimic other illnesses and the causes of the respiratory complaints are unknown [86]. For the detection of generalised TB, PCR is used to

isolate *M. tuberculosis* from sputum, body fluids, or biopsy specimens (150).

On the other hand, it was shown that 36% of the clinical indications of EPTB and 50% of the urine and/or blood samples from persons with propagating TB were PCR positive. In this investigation, the MPB-64 peptide gene was targeted by PCR to detect propagating TB in bone marrow discharges, and clinical recovery was seen in 85% of those who had positive PCR results. For the diagnosis of certain EPTB types, PCR for identifying tuberculosis (*M. tuberculosis* in blood and urine samples is a helpful tool, especially in cases when collecting the samples is challenging or invasive methods (such as tissue biopsies) are necessary. Many investigators have looked into how well PCR works to distinguish between different medical EPTB forms. 133 Oh et al. (2001) It has previously been reported that a combination of the microorganism Growth Indicator Tube (MGIT) technique, duplex PCR (multiplex PCR), targeting the 16S rRNA gene and IS6110, the CobasAmplicor System, and a "extended gold standard" made up of "true genetic material positive samples" collected from culture and medical data" can quickly identify and distinguish between *M. tuberculosis* and NTM. In a later investigation, Torrea et al. (2005) [167] devised a nested PCR approach target IS6110 for the detection of various EPTB types in urine samples from HIV-infected and non-infected people.

In Sub-Saharan African nations like Burkina Faso, there are many that have elevated HIV the prevalence of infection rates. PCR responses varied between HIV-positive and HIV-negative individuals. In detecting various EPTB kinds, two different PCR nested methods—in-house traditional PCR and LightCycler technologies addressing IS6110—were contrasted [147]. The LightCycler approach was shown to be better than the for-house approach in blood vessel aspirates, but both approaches were similarly accurate in diseased tissue specimens. Urine specimens from people with and without HIV.

In order to identify tuberculosis strains in a variety of EPTB samples (tissue tissue samples, pulmonary fluid, CSF, urine, etc.), Noussair et al., 2009 [130] established a culture-enhanced PCR method which is simultaneously responsive and particular. This test uses a broth culture in BacT/Alert MP containers as an initial stage, then proceeds to the identification of *M. TB* employing the Gene Type Mycobacterium Direct Rheumatoid arthritis (RA) and Crohn's disease patients have both had EPTB brought on by TNF- α inhibitors (such as infliximab and etanercept) and [92, 52].

GeneXpert and rifampin resistance: Rifampin is sensitivity and GeneXpert: a test Regarding EPTB clients, especially ones who have been confirmed infected with HIV resistance to drugs is a significant problem. Two significant forms of resistance to drugs are XDR-TB (Extensively Drug-Resistant Tuberculosis) and MDR-TB (Multidrug-Resistant Tuberculosis) [51]. The standard susceptibility to drugs testing needs a minimum of 2 months after the moment that the sample is contaminated. Because > 90% of RIF-isolates that are resistant also exhibit the isoniazid (INH) resistance, RIF resistance is used as an alternative biomarker for identifying MDR [62, 85]. Both the phage amplifying living test that relies on the incapacity of prone *M. tuberculosis* strains to generate RIF and a phage amplifying natural testing dependent on the lack of vulnerable *M. TB* to produce RIF had previously showed two quick physical tests for the identification of RIF opposition in tuberculosis strains. generate RIF. In

combination with of suppressive doses of RIF, tuberculosis isolates were employed to boost phage D29 reproduction. The stimulated *dnaK* (Rv0350) transcript concentrations in susceptible strains injected with RIF were shown to be lower using the RT-PCR test. In order to investigate the rapid detection of RIF resistance in *M. tuberculosis*, Brodie & Schluger (2009) [62] used line probe assays and molecular symbols real-time PCR. The recently developed Xpert test, which uses molecular beacon technology and focuses on the *rpoB* gene of wild type *M. tuberculosis* strains, has been shown to be a rapid test, giving results for both TB recognition and RIF opposition in just over two hours in a single tube [100, 168]. 547 In order to evaluate the effectiveness of the Xpert test, which has been endorsed by the WHO, to a composite standard of reference (CRS), which consists of a spread, a culture report, a list of symptoms, an ATT thereafter, and other criteria, EPTB samples were analysed.

The Xpert test has a 99.6% accuracy rate for TB identification and an 81 percent sensitivity rate. Additionally, their test successfully identified 98% of phenotypic RIF-resistant patients and 94% of phenotype RIF-susceptible patients [171]. Hillemann et al. (2011) [100] demonstrated similar positive results for TB detection in 512 EPTB samples using cultured as the gold standard. It was shown that the Xpert test performed better than either of the other two tests when the specificity for TB diagnosis in EPTB samples of the Cobas TaqMan MTB screen and the IS6110-based real-time PCR assay were compared [68, 122, 168] 1476 EPTB specimens were analysed, and the Xpert assay was determined to have 81.3% sensitive and 99.8% specific if cultured and once the gold standard was cultured and clinical evaluation. Because of its rapid turnaround (less than 2 hours), decreased contamination risks, and less operators training, this improved testing for the detection of EPTB could become more affordable in underdeveloped nations [171, 168].

Immuno-PCR (PCR Amplified Immunoassay; I-PCR): Immuno-PCR (PCR Amplified Immunoassay; I-PCR) is a unique, ultrasensitive technique to identify peptide allergens, with an accuracy improvement of at least 10³-10⁴ over a comparable ELISA. It blends the flexibility of ELISA with the accuracy of NAA by Amplification. Only molecules of DNA may be detected using PCR tests. However, the nucleic acid research may not be adequate to properly use biological information since the majority of living operations, such as EPTB illnesses, include enormous quantities of peptides along with other non-nucleic acids elements in circulating. I-PCR has been used to identify protooncogenes, cytokines that and potential viral and microbial antibodies, particularly antigens of mycobacterial species [119, 121]. ESAT-6 (Rv3875), CFP-10 (Rv3874), CFP-21 (Rv1984c), and MPT-64 (Rv1980c) are *M. tb*-specific RD1 and RD2 antigens that we used an ultrasensitive I-PCR test to identify. in the form of tissues from PTB and EPTB patients [ESAT-6 (Rv3875), CFP-10 (Rv3874)] and antibodies to these antigens. At detecting up to 0.1 fg of RD antigens, our I-PCR approach was 10⁷ times more accurate than an identical ELISA. When detecting a mixture of RD 1 and RD 2 antigens in EPTB patients, our I-PCR approach was more sensitive than an equivalent ELISA [121].

Benefits: Advantages: With rapid findings and good accuracy in diagnosing, PCR assays offer a more reliable option for finding *M. tuberculosis* in paucibacillary EPTB specimens. These investigations can help in identifying the earliest signs of EPTB and have a moderate influence on treatment of the

condition, even if they cannot completely replace the conventional AFB smear, cultured verification, or histology findings.

Demerits: Disadvantages: The utilization of relatively tiny samples and a dispersion problem of microbes in these samples may be the cause of the poorer PCR test specificity seen in certain EPTB cases as opposed to lungs samples. False-positive and false-negative results are typically linked to the use of EPTB samples in PCR experiments. However, the PCR approach fails to discriminate among acute and inactive TB [138]. It can only identify vital and ineffective *M. tuberculosis*. By PCR analysis, non-nucleic acid substances are likewise invisible.

Conclusion:

Extrapulmonary TB is still challenging to diagnose. A better understanding of the pathophysiology of TB in each organ and its imaging properties can increase the detection rate in high-risk patients. The lymph nodes are the extrapulmonary organs that are typically impacted. Necrotic lymph nodes and other organ-specific imaging abnormalities enhance the risk of extrapulmonary infection being diagnosed. Extrapulmonary TB can occur in a patient regardless of how immune they are. To summarise, a variety of processes are involved in the pathophysiology of tuberculosis. Pharmaceutically, it is difficult to target these processes because the bacteria is sneaky in its use of the immune system as a weapon. Anti-VEGF medications offer a method that has only been briefly investigated in humans (bevacizumab), focusing largely on the A isoform of VEGF. Due to the success of disseminated tuberculosis and anti-VEGF therapy using both small molecules and antibodies, clinical trials are necessary to evaluate all anti-VEGF and anti-VEGFR drugs now available in the treatment of tuberculosis.

Acknowledgement

We acknowledge the resources and financial support for the study, provided by the Department of Technology Development Transfer (TDT), Department of Science and Technology (DST), New Delhi, the Order/File No: DST/TDT/BDTD/03/2021

References:

1. World Health Organization. Global tuberculosis report 2018. https://www.who.int/tb/publications/global_report/en/. Updated February 28, 2019. Accessed February 10, 2019.
2. Peto HM, Pratt RH, Harrington TA, LoBue PA, Armstrong LR. Epidemiology of extrapulmonary tuberculosis in the United States, 1993-2006. *Clin Infect Dis* 2009;49(9):1350–1357.
3. Leeds IL, Magee MJ, Kurbatova EV, et al. Site of extrapulmonary tuberculosis is associated with HIV infection. *Clin Infect Dis* 2012;55(1):75–81.
4. Nachiappan AC, Rahbar K, Shi X, et al. Pulmonary tuberculosis: role of radiology in diagnosis and management. *RadioGraphics* 2017;37(1):52–72.

5. Handa U, Mundi I, Mohan S. Nodal tuberculosis revisited: a review. *J Infect Dev Ctries* 2012;6(1):6–12
6. Yew WW, Lee J. Pathogenesis of cervical tuberculous lymphadenitis: pathways to anatomic localization. *Tuber Lung Dis* 1995;76(3):275–276
7. Reede DL, Bergeron RT. Cervical tuberculous adenitis: CT manifestations. *Radiology* 1985;154(3):701–704.
8. Hoang JK, Vanka J, Ludwig BJ, Glastonbury CM. Evaluation of cervical lymph nodes in head and neck cancer with CT and MRI: tips, traps, and a systematic approach. *AJR Am J Roentgenol* 2013;200(1):W17–W25.
9. Ahuja A, Ying M, Yuen YH, Metreweli C. Power Doppler sonography to differentiate tuberculous cervical lymphadenopathy from nasopharyngeal carcinoma. *AJNR Am J Neuroradiol* 2001;22(4):735–740
10. Solari L, Soto A, Agapito JC, et al. The validity of cerebrospinal fluid parameters for the diagnosis of tuberculous meningitis. *Int J Infect Dis* 2013;17(12):e1111–e1115.
11. Bernaerts A, Vanhoenacker FM, Parizel PM, et al. Tuberculosis of the central nervous system: overview of neuroradiological findings. *Eur Radiol* 2003;13(8):1876–1890.
12. Mezochow A, Thakur K, Vinnard C. Tuberculous meningitis in children and adults: new insights for an ancient foe. *Curr Neurol Neurosci Rep* 2017;17(11):85.
13. Abdel Razek AA, Alvarez H, Bagg S, Refaat S, Castillo M. Imaging spectrum of CNS vasculitis. *RadioGraphics* 2014;34(4):873–894.
14. Lammie GA, Hewlett RH, Schoeman JF, Donald PR. Tuberculous cerebrovascular disease: a review. *J Infect* 2009;59(3):156–166.
15. Whiteman M, Espinoza L, Post MJ, Bell MD, Falcone S. Central nervous system tuberculosis in HIV-infected patients: clinical and radiographic findings. *AJNR Am J Neuroradiol* 1995;16(6):1319–1327.
16. Lee GT, Antelo F, Mlikotic AA. Best cases from the AFIP: cerebral toxoplasmosis. *RadioGraphics* 2009;29(4): 1200–1205.
17. Shih RY, Koeller KK. Bacterial, Fungal, and Parasitic Infections of the Central Nervous System: Radiologic-Pathologic Correlation and Historical Perspectives. *RadioGraphics* 2015;35(4):1141–1169.
18. Garg RK, Somvanshi DS. Spinal tuberculosis: a review. *J Spinal Cord Med* 2011;34(5):440–454.
19. Matos MJ, Bacelar MT, Pinto P, Ramos I. Genitourinary tuberculosis. *Eur J Radiol* 2005;55(2):181–187.
20. Merchant S, Bharati A, Merchant N. Tuberculosis of the genitourinary system: urinary tract tuberculosis—renal tuberculosis (part I). *Indian J Radiol Imaging* 2013;23(1):46–63.
21. Merchant S, Bharati A, Merchant N. Tuberculosis of the genitourinary system: urinary tract tuberculosis—renal tuberculosis (part II). *Indian J Radiol Imaging* 2013;23(1):64–77.
22. Gibson MS, Puckett ML, Shelly ME. Renal tuberculosis. *RadioGraphics* 2004;24(1):251–256.
23. Potenta SE, D’Agostino R, Sternberg KM, Tatsumi K, Perusse K. CT Urography for Evaluation of the Ureter. *RadioGraphics* 2015;35(3):709–726.

24. Jung YY, Kim JK, Cho KS. Genitourinary tuberculosis: comprehensive cross-sectional imaging. *AJR Am J Roentgenol* 2005;184(1):143–150.
25. Engin G, Acunaş B, Acunaş G, Tunaci M. Imaging of extrapulmonary tuberculosis. *RadioGraphics* 2000;20(2):471–488; quiz 529–530, 532.
26. Lakmichi MA, Kamaoui I, Eddafali B, et al. An unusual presentation of primary male genital tuberculosis. *Rev Urol* 2011;13(3):176–178.
27. Jacob JT, Nguyen TM, Ray SM. Male genital tuberculosis. *Lancet Infect Dis* 2008;8(5):335–342.
28. Li Y, Mongan J, Behr SC, et al. Beyond prostate adenocarcinoma: expanding the differential diagnosis in prostate pathologic conditions. *RadioGraphics* 2016;36(4):1055–1075.
29. Sharma JB. Current diagnosis and management of female genital tuberculosis. *J ObstetGynaecol India* 2015;65(6):362–371.
30. Grace GA, Devaleenal DB, Natrajan M. Genital tuberculosis in females. *Indian J Med Res* 2017;145(4):425–436.
31. Cho JK, Choi YM, Lee SS, et al. Clinical features and outcomes of abdominal tuberculosis in southeastern Korea: 12 years of experience. *BMC Infect Dis* 2018;18(1):699.
32. Pereira JM, Madureira AJ, Vieira A, Ramos I. Abdominal tuberculosis: imaging features. *Eur J Radiol* 2005;55(2):173–180.
33. Sharma R, Madhusudhan KS, Ahuja V. Intestinal tuberculosis versus Crohn's disease: clinical and radiological recommendations. *Indian J Radiol Imaging* 2016;26(2):161–172.
34. Burrill J, Williams CJ, Bain G, Conder G, Hine AL, Misra RR. Tuberculosis: a radiologic review. *RadioGraphics* 2007;27(5):1255–1273.
35. Bächler P, Baladron MJ, Menias C, et al. Multimodality imaging of liver infections: differential diagnosis and potential pitfalls. *RadioGraphics* 2016;36(4):1001–1023.
36. Sargar KM, Khanna G, Hulett Bowling R. Imaging of nonmalignant adrenal lesions in children. *RadioGraphics* 2017;37(6):1648–1664.
37. Almeida A. Tuberculosis of the spine and spinal cord. *Eur J Radiol* 2005;55(2):193–201.
38. Kamara E, Mehta S, Brust JC, Jain AK. Effect of delayed diagnosis on severity of Pott's disease. *Int Orthop* 2012;36(2):245–254.
39. Kilborn T, Janse van Rensburg P, Candy S. Pediatric and adult spinal tuberculosis: imaging and pathophysiology. *Neuroimaging Clin N Am* 2015;25(2):209–231.
40. Rajasekaran S, Soundararajan DCR, Shetty AP, Kanna RM. Spinal tuberculosis: current concepts. *Global Spine J* 2018;8(4 suppl):96S–108S.
41. Jung NY, Jee WH, Ha KY, Park CK, Byun JY. Discrimination of tuberculous spondylitis from pyogenic spondylitis on MRI. *AJR Am J Roentgenol* 2004;182(6):1405–1410.
42. Harada Y, Tokuda O, Matsunaga N. Magnetic resonance imaging characteristics of tuberculous spondylitis vs. pyogenic spondylitis. *Clin Imaging* 2008;32(4):303–309.
43. Parmar H, Shah J, Patkar D, Singrakhia M, Patankar T, Hutchinson C. Tuberculous arthritis of the appendicular skeleton: MR imaging appearances. *Eur J Radiol* 2004;52(3):300–309.
44. De Backer AI, Mortelé KJ, Vanhoenacker FM, Parizel PM. Imaging of extraspinal musculoskeletal tuberculosis. *Eur J Radiol* 2006;57(1):119–130.

45. Sawlani V, Chandra T, Mishra RN, Aggarwal A, Jain UK, Gujral RB. MRI features of tuberculosis of peripheral joints. *Clin Radiol* 2003;58(10):755–762.
46. Sanghvi DA, Iyer VR, Deshmukh T, Hoskote SS. MRI features of tuberculosis of the knee. *Skeletal Radiol* 2009;38(3):267–273.
47. Wells D, Strickland C, Schowinsky J, Lindeque B. Nontuberculous mycobacterial tenosynovitis: AIRP best cases in radiologic-pathologic correlation. *RadioGraphics* 2015;35(2):493–497.
48. Dias MF, Bernardes Filho F, Quaresma MV, Nascimento LV, Nery JA, Azulay DR. Update on cutaneous tuberculosis. *An Bras Dermatol* 2014;89(6):925–938.
49. Abbara A & Davidson RN (2011) Etiology and management of genitourinary tuberculosis. *Nat Rev Urol* 8: 678–688.
50. Abdalla CM, de Oliveira ZN, Sotto MN, Leite KR, Canavez FC & de Carvalho CM (2009) Polymerase chain reaction compared to other laboratory findings and to clinical evaluation in the diagnosis of cutaneous tuberculosis and atypical mycobacteria skin infection. *Int J Dermatol* 48: 27–35.
51. Agashe V, Shenai S, Mohrir G, Deshmukh M, Bhaduri A, Deshpande R, Mehta A & Rodrigues C (2009) Osteoarticular tuberculosis-diagnostic solutions in a disease endemic region. *J Infect Dev Ctries* 3: 511–516.
52. Almadi MA, Ghosh S & Aljebreen AM (2009) Differentiating intestinal tuberculosis from Crohn's disease: a diagnostic challenge. *Am J Gastroenterol* 104: 1003–1012.
53. Alvarez GG, Roth VR & Hodge W (2009) Ocular tuberculosis: diagnostic and treatment challenges. *Int J Infect Dis* 13: 432–435.
54. Baba K, Pathak S, Sviland L, Langeland N, Hoosen AA, Asjo B, Dyrhol-Riise AM & Mustafa T (2008) Real-time quantitative PCR in the diagnosis of tuberculosis in formalin-fixed paraffin-embedded pleural tissue in patients from a high HIV endemic area. *Diagn Mol Pathol* 17: 112–117.
55. Baek CH, Kim SI, Ko YH & Chu KC (2000) Polymerase chain reaction detection of *Mycobacterium tuberculosis* from fineneedle aspirate for the diagnosis of cervical tuberculous lymphadenitis. *Laryngoscope* 110: 30–34.
56. Baharoon S (2008) Tuberculosis of the breast. *Ann Thorac Med* 3: 110–114.
57. Balamurugan R, Chittaranjan S, Subramanian V & Ramakrishna BS (2010) Fecal polymerase chain reaction for *Mycobacterium tuberculosis* IS6110 to distinguish Crohn's disease from intestinal tuberculosis. *Indian J Gastroenterol* 29: 152–156.
58. Bandyopadhyay D, Gupta S, Banerjee S, Gupta S, Ray D, Bhattacharya S & Bhattacharya B (2008) Adenosine deaminase estimation and multiplex polymerase chain reaction in diagnosis of extra-pulmonary tuberculosis. *Int J Tuberc Lung Dis* 12: 1203–1208.
59. Bhanu NV, Singh UB, Chakraborty M, Suresh N, Arora J, Rana T, Takkar D & Seth P (2005) Improved diagnostic value of PCR in the diagnosis of female genital tuberculosis leading to infertility. *J Med Microbiol* 54: 927–931.

60. Boëddinghaus B, Wichelhaus TA, Brade V & Bittner T (2001) Removal of PCR inhibitors by silica membranes: evaluating the Amplicor Mycobacterium tuberculosis kit. *J Clin Microbiol* 39: 3750–3752.
61. Bravo FG & Gotuzzo E (2007) Cutaneous tuberculosis. *Clin Dermatol* 25: 173–180.
62. Brodie D & Schluger NW (2009) Nucleic acid amplification for detection of Mycobacterium tuberculosis. *Tuberculosis: A Comprehensive Clinical Reference* (Schaaf HS & Zumla A, eds), pp. 197–204.
63. Saunders Elsevier, Oxford. Bruijnesteijn Van Coppenraet ES, Lindeboom JA, Prins JM, Peeters MF, Claas ECJ & Kuijper EJ (2004) Real-time PCR assay using fine needle aspirates and tissue biopsy specimens for rapid diagnosis of mycobacterial lymphadenitis in children. *J Clin Microbiol* 42: 2644–2650.
64. Bulbuloglu E, Ciralik H, Okur E, Ozdemir G, Ezberci F & Cetinkaya A (2006) Tuberculosis of the thyroid gland: review of the literature. *World J Surg* 30: 149–155.
65. Burgess LJ, Reuter H, Carstens ME, Taljaard JJ & Doubell AF (2002) The use of adenosine deaminase and interferon gamma as diagnostic tools for tuberculous pericarditis. *Chest* 122: 900–905.
66. Cabandugama P, Pores N & Feldstein RC (2011) Abdominal tuberculosis. *Pract Gastroenterol* 35: 38–44.
67. Catedral EJ, Santos RE, Padilla MDB & Fajardo-Ang C (2010) Detection of Mycobacterium tuberculosis in corneas from donors with active tuberculosis disease through polymerase chain reaction and culture. *Br J Ophthalmol* 94: 894–897.
68. Causse M, Ruiz P, Gutierrez-Aroca JB & Casal M (2011) Comparison of two molecular methods for rapid diagnosis of extrapulmonary tuberculosis. *J Clin Microbiol* 49: 3065–3067.
69. Chakravorty S & Tyagi JS (2005) Novel multipurpose methodology for detection of mycobacteria in pulmonary and extrapulmonary specimens by smear microscopy, culture, and PCR. *J Clin Microbiol* 43: 2697–2702.
70. Chakravorty S, Sen MK & Tyagi JS (2005) Diagnosis of extrapulmonary tuberculosis by smear, culture, and PCR using universal sample processing technology. *J Clin Microbiol* 43: 4357–4362.
71. Chawla K, Gupta S, Mukhopadhyay C, Rao PS & Bhat SS (2009) PCR for M. tuberculosis in tissue samples. *J Infect Dev Ctries* 3: 83–87.
72. Cheng VC, Yam WC, Hung IF, Woo PC, Lau SK, Tang BS & Yuen KY (2004) Clinical evaluation of the polymerase chain reaction for the rapid diagnosis of tuberculosis. *J Clin Pathol* 57: 281–285.
73. Cheng VC, Yew WW & Yuen KY (2005) Molecular diagnostics in tuberculosis. *Eur J Clin Microbiol Infect Dis* 24: 711–720.
74. Cherian G (2004) Diagnosis of tuberculous aetiology in pericardial effusions. *Postgrad Med J* 80: 262–266.
75. Choi SR, Kim JK, Kim DH & Yoon MS (2009) A case of tuberculosis cutis orificialis with perianal involvement. *Ann Dermatol* 21: 443–446.

76. Colmenero JD, Morata P, Ruiz-Mesa JD, Bautista D, Bermudez P, Bravo MJ & Queipo-Ortun˜o MI (2010) Multiplex real-time polymerase chain reaction: a practical approach for rapid diagnosis of tuberculous and brucellar vertebral osteomyelitis. *Spine* 35: E1392–E1396.
77. Cortez MV, Oliveira CM, Monte RL, Arauˆjo JR, Braga BB, Reis DZ, Ferreira LC, Moraes MO & Talhari S (2011) HIV-associated tuberculous lymphadenitis: the importance of polymerase chain reaction (PCR) as a complementary tool for the diagnosis of tuberculosis – a study of 104 patients. *An Bras Dermatol* 86: 925–931.
78. Cutrufello NJ, Karakousis PC, Fishler J & Albini TA (2010) Intraocular tuberculosis. *Ocul Immunol Inflamm* 18: 281–291.
79. Dale JW, Al-Ghusein H, Al-Hashmi S et al. (2003) Evolutionary relationships among strains of *Mycobacterium tuberculosis* with few copies of IS6110. *J Bacteriol* 185: 2555–2562.
80. Daley P, Thomas S & Pai M (2007) Nucleic acid amplification tests for the diagnosis of tuberculous lymphadenitis: a systematic review. *Int J Tuberc Lung Dis* 11: 1166–1176.
81. Derese Y, Hailu E, Assefa T, Bekele Y, Mihret A, Aseffa A, Hussien J, Ali I & Abebe M (2012) Comparison of PCR with standard culture of fine needle aspiration samples in the diagnosis of tuberculosis lymphadenitis. *J Infect Dev Ctries* 6: 53–57.
82. Desai D, Nataraj G, Kulkarni S, Bichile L, Mehta P, Baveja S, Rajan R, Raut A & Shenoy A (2006) Utility of the polymerase chain reaction in the diagnosis of tuberculous meningitis. *Res Microbiol* 157: 967–970.
83. Dora JM, Geib G, Chakr R, Paris F, Mombach AB, Lutz L, Souza CF & Goldani LZ (2008) Polymerase chain reaction as a useful and simple tool for rapid diagnosis of tuberculous meningitis in a Brazilian tertiary care hospital. *Braz J Infect Dis* 12: 245–247.
84. Dyrhol-Riise AM, Gran G, Wentzel-Larsen T, Blomberg B, Haanshuus CG & Mørkve O (2010) Diagnosis and follow-up of treatment of latent tuberculosis; the utility of the QuantiFERON-TB Gold In-tube assay in outpatients from a tuberculosis low-endemic country. *BMC Infect Dis* 10: 57.
85. Eltringham IJ, Drobniewski FA, Mangan JA, Butcher PD & Wilson SM (1999) Evaluation of reverse transcription-PCR and a bacteriophage-based assay for rapid phenotypic detection of rifampin resistance in clinical isolates of *Mycobacterium tuberculosis*. *J Clin Microbiol* 37: 3524–3527.
86. Escobedo-Jaimes L, Cicero-Sabido R, Criales-Cortez JL, Ramirez E, Romero M, Rivero V, Islas F, Olivera H, Gonzalez S & Escobar-Gutierrez A (2003) Evaluation of the polymerase chain reaction in the diagnosis of miliary tuberculosis in bone marrow smear. *Int J Tuberc Lung Dis* 7: 580–586.
87. Fujimoto N, Gemba K, Yao A, Ozaki S, Ono K, Wada S, Fujii Y, Namba Y & Kishimoto T (2010) Tuberculosis diagnosed by PCR analysis of synovial fluid. *J Infect Chemother* 16: 53–55.
88. Galimi R (2011) Extrapulmonary tuberculosis: tuberculous meningitis new developments. *Eur Rev Med Pharmacol Sci* 15: 365–386.

89. Gan HT, Chen YQ, Ouyang Q, Bu H & Yang XY (2002) Differentiation between intestinal tuberculosis and Crohn's disease in endoscopic biopsy specimens by polymerase chain reaction. *Am J Gastroenterol* 97: 1446–1451.
90. Garcia-Elorriaga G, Gracida-Osorno C, Carrillo-Montes G & Gonzalez-Bonilla C (2009) Clinical usefulness of the nested polymerase chain reaction in the diagnosis of extrapulmonary tuberculosis. *Salud Publica Mex* 51: 240–245.
91. Ghosh A, Saha S, Bhattacharya B & Chattopadhyay S (2007) Primary tuberculosis of thyroid gland: a rare case report. *Am J Otolaryngol* 28: 267–270.
92. Golden MP & Vikram HR (2005) Extrapulmonary tuberculosis: an overview. *Am Fam Physician* 72: 1761–1768.
93. Green C, Huggett JF, Talbot E, Mwaba P, Reither K & Zumla AI (2009) Rapid diagnosis of tuberculosis through the detection of mycobacterial DNA in urine by nucleic acid amplification methods. *Lancet Infect Dis* 9: 505–511.
94. Griffiths G, Nystrom B, Sable SB & Khuller GK (2010) Nanobead-based interventions for the treatment and prevention of tuberculosis. *Nat Rev Microbiol* 8: 827–834.
95. Gupta V, Gupta A & Rao NA (2007) Intraocular tuberculosis: an update. *Surv Ophthalmol* 52: 561–587.
96. Haldar S, Sharma N, Gupta VK & Tyagi JS (2009) Efficient diagnosis of tuberculous meningitis by detection of *Mycobacterium tuberculosis* DNA in cerebrospinal fluid filtrates using PCR. *J Med Microbiol* 58: 616–624.
97. Haldar S, Bose M, Chakrabarti P et al. (2011) Improved laboratory diagnosis of tuberculosis – the Indian experience. *Tuberculosis* 91: 414–426.
98. Harari A, Rozot V, Enders FB et al. (2011) Dominant TNF- α *Mycobacterium tuberculosis*-specific CD4 + T cell responses discriminate between latent infection and active disease. *Nat Med* 17: 372–376.
99. Hemal AK, Gupta NP, Rajeev TP, Kumar R, Dar L & Seth P (2000) Polymerase chain reaction in clinically suspected genitourinary tuberculosis: comparison with intravenous urography, bladder biopsy, and urine acid fast bacilli culture. *Urology* 56: 570–574.
100. Hillemann D, Rusch-Gerdes S, Boehme C & Richter E (2011) Rapid molecular detection of extrapulmonary tuberculosis by the automated GeneXpert MTB/RIF system. *J Clin Microbiol* 49: 1202–1205.
101. Honore-Bouakline S, Vincensini JP, Giacuzzo V, Lagrange PH & Herrmann JL (2003) Rapid diagnosis of extrapulmonary tuberculosis by PCR: impact of sample preparation and DNA extraction. *J Clin Microbiol* 41: 2323–2329.
102. Jain AK, Jena SK, Singh MP, Dhammi IK, Ramachandran VG & Dev G (2008) Evaluation of clinico-radiological, bacteriological, serological, molecular and histological diagnosis of osteoarticular tuberculosis. *Indian J Orthop* 42: 173–177.
103. Jambhekar NA, Kulkarni SP, Madur BP, Agarwal S & Rajan MG (2006) Application of the polymerase chain reaction on formalin-fixed, paraffin-embedded tissue in the recognition of tuberculous osteomyelitis. *J Bone Joint Surg Br* 88: 1097–1101.

104. Jin XJ, Kim JM, Kim HK et al. (2010) Histopathology and TBPCR kit analysis in differentiating the diagnosis of intestinal tuberculosis and Crohn's disease. *World J Gastroenterol* 16: 2496–2503.
105. Kalantri Y, Hemvani N & Chitnis DS (2011) Evaluation of real-time polymerase chain reaction, interferon-gamma, adenosine deaminase, and immunoglobulin A for the efficient diagnosis of pleural tuberculosis. *Int J Infect Dis* 15: e226–e231.
106. Kalra M, Khuller GK, Grover A, Behera D, Wanchu A & Verma I (2010) Utility of a combination of RD1 and RD2 antigens as a diagnostic marker for tuberculosis. *Diagn Microbiol Infect Dis* 66: 153–161.
107. Kao PT, Tu MY, Tang SH & Ma HK (2010) Tuberculosis of the breast with erythema nodosum: a case report. *J Med Case Rep* 29: 124.
108. Katoch VM (2004) Newer diagnostic techniques for tuberculosis. *Indian J Med Res* 120: 418–428.
109. Kulkarni SP, Jaleel MA & Kadival GV (2005) Evaluation of an in-house developed PCR for the diagnosis of tuberculous meningitis in Indian children. *J Med Microbiol* 54: 369–373.
110. Kulkarni S, Vyas S, Supe A & Kadival G (2006) Use of polymerase chain reaction in the diagnosis of abdominal tuberculosis. *J Gastroenterol Hepatol* 21: 819–823.
111. Kulkarni S, Rajan MGR, Hazra P & Islam A (2011) Development of a TB-PCR kit for the diagnosis of tuberculosis. *BARC Newsletter* 319: 44–50.
112. Lange C & Mori T (2010) Advances in the diagnosis of tuberculosis. *Respirology* 15: 220–240.
113. Laniado-Laborin R (2005) Adenosine deaminase in the diagnosis of tuberculous pleural effusion: is it really an ideal test? A word of caution. *Chest* 127: 417–418.
114. Light RW (2010) Update on tuberculous pleural effusion. *Respirology* 15: 451–458.
115. Lima DM, Colares JK & da Fonseca BA (2003) Combined use of the polymerase chain reaction and detection of adenosine deaminase activity on pleural fluid improves the rate of diagnosis of pleural tuberculosis. *Chest* 124: 909–914.
116. KT, Su WJ & Perng RP (2007) Clinical utility of polymerase chain reaction for diagnosis of smear-negative pleural tuberculosis. *J Chin Med Assoc* 70: 148–151.
117. Madhavan HN, Therese KL, Gunisha P, Jayanthi U & Biswas J (2000) Polymerase chain reaction for detection of *Mycobacterium tuberculosis* in epiretinal membrane in Eales' disease. *Invest Ophthalmol Vis Sci* 41: 822–825.
118. Madhavan HN, Therese KL & Doraiswamy K (2002) Further investigations on the association of *Mycobacterium tuberculosis* with Eales' disease. *Indian J Ophthalmol* 50: 35–39.
119. Malou N & Raoult D (2011) Immuno-PCR: a promising ultrasensitive diagnostic method to detect antigens and antibodies. *Trends Microbiol* 19: 295–302.
120. Martins LC, Paschoal IA, Von Nowakowski A, Silva SA, Costa FF & Ward LS (2000) Nested-PCR using MPB64 fragment improves the diagnosis of pleural and meningeal tuberculosis. *Rev Soc Bras Med Trop* 33: 253–257.
121. Mehta PK, Kalra M, Khuller GK, Behera D & Verma I (2012) Development of an ultrasensitive polymerase chain reaction amplified immunoassay (Immuno-PCR) based on

- mycobacterial RD antigens: implications for the serodiagnosis of tuberculosis. *Diagn Microbiol Infect Dis* 72: 166–174.
122. Miller MB, Popowitch EB, Backlund MG & Ager EP (2011) Performance of Xpert MTB/RIF RUO assay and IS6110 realtime PCR for *Mycobacterium tuberculosis* detection in clinical samples. *J Clin Microbiol* 49: 3458–3462.
123. Mirza S, Restrepo BI, McCormick JB & Fisher-Hoch SP (2003) Diagnosis of tuberculosis lymphadenitis using a polymerase chain reaction on peripheral blood mononuclear cells. *Am J Trop Med Hyg* 69: 461–465.
124. Mohapatra PR & Janmeja AK (2009) Tuberculous lymphadenitis. *J Assoc Physicians India* 57: 585–590.
125. Morris K (2011) WHO recommends against inaccurate tuberculosis tests. *Lancet* 377: 113–114.
126. Moussa OM, Eraky I, El-Far MA, Osman HG & Ghoneim MA (2000) Rapid diagnosis of genitourinary tuberculosis by polymerase chain reaction and non-radioactive DNA hybridization. *J Urol* 164: 584–588.
127. Negi SS, Basir SF, Gupta S, Pasha ST, Khare S & Lal S (2005a) Comparative study of PCR, smear examination and culture for diagnosis of cutaneous tuberculosis. *J Commun Dis* 37: 83–92.
128. Negi SS, Gupta S, Khare S & Lal S (2005b) Comparison of various microbiological tests including polymerase chain reaction for the diagnosis of osteoarticular tuberculosis. *Indian J Med Microbiol* 23: 245–248.
129. Nopvichai C, Sanpavat A, Sawatdee R, Assanasen T, Wacharapluesadee S, Thorner PS & Shuangshoti S (2009) PCR detection of *Mycobacterium tuberculosis* in necrotizing non-granulomatous lymphadenitis using formalin-fixed paraffin-embedded tissue: a study in Thai patients. *J Clin Pathol* 62: 812–815.
130. Noussair L, Bert F, Leflon-Guibout V, Gayet N & Nicolas-Chanoine MH (2009) Early diagnosis of extrapulmonary tuberculosis by a new procedure combining broth culture and PCR. *J Clin Microbiol* 47: 1452–1457.
131. Obieta MP, Obieta MY & Monje VD (2010) Polymerase chain reaction-based detection of *Mycobacterium tuberculosis* DNA in formalin-fixed, paraffin-embedded skin tissue specimens from Filipino patients. *Int J Dermatol* 49: 470–472.
132. Ogusku MM, Sadahiro A, Hirata MH, Hirata RDC, Zaitz C & Salem JI (2003) PCR in the diagnosis of cutaneous tuberculosis. *Braz J Microbiol* 34: 165–170.
133. Oh EJ, Park YJ, Chang CL, Kim BK & Kim SM (2001) Improved detection and differentiation of mycobacteria with combination of *Mycobacterium* Growth Indicator Tube and Roche COBAS AMPLICOR System in conjunction with Duplex PCR. *J Microbiol Methods* 46: 29–36.
134. Okazaki T, Ebihara S, Takahashi H, Asada M, Sato A, Seki M, Ohto H & Sasaki H (2005) Multiplex PCR-identified cutaneous tuberculosis evoked by *Mycobacterium bovis* BCG vaccination in a healthy baby. *J Clin Microbiol* 43: 523–525.

135. Osore F, Nolasco O, Verdonck K, Are'valo J, Ferrufino JC, Agapito J, Huayanay L, Gotuzzo E & Maguin~a C (2006) Clinical evaluation of a 16S ribosomal RNA polymerase chain reaction test for the diagnosis of lymph node tuberculosis. *Clin Infect Dis* 43: 855–859.
136. Padmavathy L, Rao L & Veliath A (2003) Utility of polymerase chain reaction as a diagnostic tool in cutaneous tuberculosis. *Indian J Dermatol Venereol Leprol* 69: 214–216.
137. Pahwa R, Hedau S, Jain S, Jain N, Arora VM, Kumar N & Das BC (2005) Assessment of possible tuberculous lymphadenopathy by PCR compared to nonmolecular methods. *J Med Microbiol* 54: 873–878.
138. Pai M & O'Brien R (2008) New diagnostics for latent and active tuberculosis: state of the art and future prospects. *Semin Respir Crit Care Med* 29: 560–568.
139. Pai M, Flores LL, Hubbard A, Riley LW & Colford JM Jr (2004) Nucleic acid amplification tests in the diagnosis of tuberculous pleuritis: a systematic review and meta-analysis. *BMC Infect Dis* 4: 6.
140. Parrish NM & Carroll KC (2011) Role of the clinical mycobacteriology laboratory in diagnosis and management of tuberculosis in low-prevalence settings. *J Clin Microbiol* 49: 772–776.
141. Peto HM, Pratt RH, Harrington TA, LoBue PA & Armstrong LR (2009) Epidemiology of extrapulmonary tuberculosis in the United States, 1993-2006.
142. *Clin Infect Dis* 49: 1350–1357. Pulimood AB, Peter S, Rook GW & Donoghue HD (2008) In situ PCR for *Mycobacterium tuberculosis* in endoscopic mucosal biopsy specimens of intestinal tuberculosis and Crohn disease. *Am J Clin Pathol* 129: 846–851.
143. Quan C, Lu CZ, Qiao J, Xiao BG & Li X (2006) Comparative evaluation of early diagnosis of tuberculous meningitis by different assays. *J Clin Microbiol* 44: 3160–3166.
144. Rafi W, Venkataswamy MM, Ravi V & Chandramuki A (2007) Rapid diagnosis of tuberculous meningitis: a comparative evaluation of in-house PCR assays involving three mycobacterial DNA sequences, IS6110, MPB-64 and 65 kDa antigen. *J Neurol Sci* 252: 163–168.
145. Rana T, Singh UB, Kulshrestha V, Kaushik A, Porwal C, Agarwal N & Kriplani A (2011) Utility of reverse transcriptase PCR and DNA-PCR in the diagnosis of female genital tuberculosis. *J Med Microbiol* 60: 486–491.
146. Rebollo MJ, San Juan Garrido R, Folgueira D, Palenque E, Di'az-Pedroche C, Lumbreras C & Aguado JM (2006) Blood and urine samples as useful sources for the direct detection of tuberculosis by polymerase chain reaction. *Diagn Microbiol Infect Dis* 56: 141–146.
147. Ritis K, Giaglis S, Rafail S, Alepopoulou E, Tsironidou V, Tzoanopoulos D, Speletas M, Ktenidou-Kartali S, Sideras P & Kartalis G (2005) Diagnostic usefulness of bone marrow aspiration material for the amplification of IS6110 insertion element in extrapulmonary tuberculosis: comparison of two PCR techniques. *Int J Tuberc Lung Dis* 9: 455–460.
148. Rosso F, Michelon CT, Sperhackle RD, Verza M, Olival L, Conde MB, Guerra RL, Zaha A & Rossetti ML (2011) Evaluation of real-time PCR of patient pleural effusion for diagnosis of tuberculosis. *BMC Res Notes* 4: 279.

149. Santos A, Cremades R, Rodríguez JC, García-Pachón E, Ruiz M & Royo G (2009) Comparison of methods of DNA extraction for real-time PCR in a model of pleural tuberculosis. *APMIS* 118: 60–65.
150. Sharma SK & Banga A (2005) Pleural fluid interferon-gamma and adenosine deaminase levels in tuberculosis pleural effusion: a cost-effectiveness analysis. *J Clin Lab Anal* 19: 40–46.
151. Sharma SK & Mohan A (2004) Extrapulmonary tuberculosis. *Indian J Med Res* 120: 316–353.
152. Sharma SK, Mohan A, Sharma A & Mitra DK (2005) Miliary tuberculosis: new insights into an old disease. *Lancet Infect Dis* 5: 415–430.
153. Sharma K, Sharma A, Singh M, Ray P, Dandora R, Sharma SK, Modi M, Prabhakar S & Sharma M (2010a) Evaluation of polymerase chain reaction using protein b primers for rapid diagnosis of tuberculous meningitis. *Neurol India* 58: 727–731.
154. Sharma M, Sethi S, Mishra AK, Chatterjee SS, Wanchu A & Nijhawan R (2010b) Efficacy of an in-house polymerase chain reaction assay for rapid diagnosis of *Mycobacterium tuberculosis* in patients with tubercular lymphadenitis: comparison with fine needle aspiration cytology and conventional techniques. *Indian J Pathol Microbiol* 53: 714–717.
155. Sharma K, Sharma A, Ray P, Sharma SK, Modi M, Prabhakar S, Varma S & Sharma M (2011a) Multiplex PCR for rapid diagnosis of tuberculous meningitis. *J Neurol* 258: 1781–1787.
156. Sharma K, Sharma A, Sharma SK, Sen RK, Dhillon MS & Sharma M (2011b) Does multiplex polymerase chain reaction increase the diagnostic percentage in osteoarticular tuberculosis? A prospective evaluation of 80 cases. *Int Orthop* 36: 255–259.
157. Sharma P, Bansal R, Gupta V & Gupta A (2011c) Diagnosis of tubercular uveitis by quantitative polymerase chain reaction. *J Ophthalmic Inflamm Infect* 1: 23–27.
158. Singh UB, Bhanu NV, Suresh VN, Arora J, Rana T & Seth P (2006) Utility of polymerase chain reaction in diagnosis of tuberculosis from samples of bone marrow aspirate. *Am J Trop Med Hyg* 75: 960–963.
159. Srivastava R, Kumar D, Waskar MN, Sharma M, Katoh VM & Srivastava BS (2006) Identification of a repetitive sequence belonging to a PPE gene of *Mycobacterium tuberculosis* and its use in diagnosis of tuberculosis. *J Med Microbiol* 55: 1071–1077.
160. Steingart KR, Flores LL, Dendukuri N, Schiller I, Laal S, Ramsay A, Hopewell PC & Pai M (2011) Commercial serological tests for the diagnosis of active pulmonary and extrapulmonary tuberculosis: an updated systematic review and meta-analysis. *PloS Med* 8: e1001062.
161. Sun L, Yuan Q, Feng JM, Yang CM, Yao L, Fan QL, Liu LL, Ma JF & Wang LN (2010) Rapid diagnosis in early stage renal tuberculosis by real-time polymerase chain reaction on renal biopsy specimens. *Int J Tuberc Lung Dis* 14: 341–346.
162. Sun YS, Lou SQ, Wen JM, Lv WX, Jiao CG, Yang SM & Xu HB (2011) Clinical value of polymerase chain reaction in the diagnosis of joint tuberculosis by detecting the DNA of *Mycobacterium tuberculosis*. *Orthop Surg* 3: 64–71.
163. Supiyaphun P, Tumwasornb S, Udomsantisukb N, Keelawatc S, Songsrisangaa W, Prasurthsinb P & Sawatpanichb A (2010) Diagnostic tests for tuberculous lymphadenitis: fine

- needle aspirations using tissue culture in mycobacteria growth indicator tube and tissue PCR. *Asian Biomed* 4: 787–792.
164. Sutherland JS, Garba D, Fombah AE, Mendy-Gomez A, Mendy FS, Antonio M, Townend J, Ideh RC, Corrah T & Ota MO (2012) Highly accurate diagnosis of pleural tuberculosis by immunological analysis of the pleural effusion. *PLoS ONE* 7: e30324.
 165. Takahashi T, Tamura M, Asami Y et al. (2008) Novel wide-range quantitative nested real-time PCR assay for *Mycobacterium tuberculosis* DNA: clinical application for diagnosis of tuberculous meningitis. *J Clin Microbiol* 46: 1698–1707.
 166. Thangappah RB, Paramasivan CN & Narayanan S (2011) Evaluating PCR, culture & histopathology in the diagnosis of female genital tuberculosis. *Indian J Med Res* 134: 40–46.
 167. Torrea G, Van de Perre P, Ouedraogo M et al. (2005) PCRbased detection of the *Mycobacterium tuberculosis* complex in urine of HIV-infected and uninfected pulmonary and extrapulmonary tuberculosis patients in Burkina Faso. *J Med Microbiol* 54: 39–44.
 168. Tortoli E, Russo C, Piersimoni C et al. (2012) Clinical validation of Xpert MTB/RIF for the diagnosis of extrapulmonary tuberculosis. *Eur Respir J*. DOI: 10.1183/09031936.00176311.
 169. Totsch M, Bocker W, Brommelkamp E, Fille M, Kreczy A, Ofner D, Schmid KW & Dockhorn-Dworniczak B (1996) Diagnostic value of different PCR assays for the detection of mycobacterial DNA in granulomatous lymphadenopathy. *J Pathol* 178: 221–226.
 170. Tzoanopoulos D, Stakos D, Hatseras D, Ritis K & Kartalis G (2001) Detection of *Mycobacterium tuberculosis* complex DNA in pericardial fluid, bone marrow and peripheral blood in a patient with pericardial tuberculosis. A case report. *Neth J Med* 59: 177–180.
 171. KR, Talluri Rameshwari & Khan, Nazia & Malgotra, Vikas & Kashwani, Ritik & Shanmugam, Arun & Kumar, Sumana. (2024). Histopathological studies and Cellular changes of *Mycobacterium tuberculosis* in Extra Pulmonary Tuberculosis in tertiary care Hospital in India. *African Journal of Biological Sciences*. 6. 308-320. 10.33472/AFJBS.6.5.2024
 172. Vardhan V & Yanamandra U (2011) Diagnosis of osteoarticular tuberculosis. *Indian J Rheumatol* 6: 87–94.
 173. Verettas D, Kazakos C, Tilkeridis C, Dermon A, Petrou H & Galanis V (2003) Polymerase chain reaction for the detection of *Mycobacterium tuberculosis* in synovial fluid, tissue samples, bone marrow aspirate and peripheral blood. *Acta Orthop Belg* 69: 396–399.
 174. Verma P, Jain A, Patra SK, Gandhi S, Sherwal BL & Chaudhary M (2010) Evaluation of polymerase chain reaction (PCR) using hupB gene in diagnosis of tuberculous lymphadenitis in fine needle aspirates. *Indian J Tuberc* 57: 128–133.
 175. Villegas MV, Labrada LA & Saravia NG (2000) Evaluation of polymerase chain reaction, adenosine deaminase, and interferon-gamma in pleural fluid for the differential diagnosis of pleural tuberculosis. *Chest* 118: 1355–1364.
 176. Wise GJ (2009) Urinary tuberculosis: modern issues. *Curr Urol Rep* 10: 313–318.
 177. WHO (2011) Global tuberculosis control. WHO report. World Health Organization, Geneva.
 178. Vandana, Kr, Talluri & Khan, Nazia & Bhat, Ramdas & Sagili, Sreenivas & Kashwani, Ritik & Aman, Awanish & Kumar, Sumana. (2024). MYCOBACTERIUM TUBERCULOSIS

- H37Rv GENE EXPRESSION OF THE OmpA FAMILY PROTEINS. *Community practitioner: the journal of the Community Practitioners' & Health Visitors' Association*. 21. 1853-1866. 10.5281/zenodo.12272526.
179. Yun YJ, Lee KH, Haihua L et al. (2005) Detection and identification of *Mycobacterium tuberculosis* in joint biopsy specimens by rpoB PCR cloning and sequencing. *J Clin Microbiol* 43: 174–178.
180. Zamirian M, Mokhtarian M, Motazedian MH, Monabati A & Reza Rezaian G (2007) Constrictive pericarditis: detection of *Mycobacterium tuberculosis* in paraffin-embedded pericardial tissues by polymerase chain reaction. *Clin Biochem* 40: 355–358